

Original Article

**Association between *ApoE* Polymorphism and Type 2 Diabetes: A Meta-Analysis of 59 Studies***CHEN Da Wei¹, SHI Ji Kang², LI Yun³, YANG Yu⁴, and REN Shu Ping^{5,#}

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Abstract

Objective To identify the important risk factors for type 2 Diabetes Mellitus (T2DM) and develop effective strategies to address the problem of T2DM. Our study aimed to evaluate the association between apolipoprotein E (*ApoE*) genetic polymorphism and type 2 diabetes, and to provide clues for the etiology of T2DM.

Methods Based on the criteria of inclusion and exclusion, we extracted, pooled, analyzed and assessed the case-control studies of *ApoE* polymorphism and T2DM published in PubMed, Web of Science, Medline, WanFang, VIP, and CNKI databases by R soft-ware (version 3.4.3). We used Random-effect models when heterogeneity was present in between-study, and fixed-effect models otherwise.

Results We had 59 studies covering 6,872 cases with T2DM and 8,250 controls, and compared the alleles and genotypes of *ApoE* between cases and controls. When we conducted a comparison between *ApoE* $\epsilon 4$ and $\epsilon 3$ alleles, we produced a pooled *OR* of 1.18 (95% *CI*: 1.09-1.28; *P* < 0.001). *ApoE* $\epsilon 2/\epsilon 2$ genotype displayed a possible association with T2DM (*OR* = 1.46; 95% *CI*: 1.11-1.93; *P* = 0.007), $\epsilon 3/\epsilon 4$ genotype showed a 1.11-fold risk (*OR* = 1.11; 95% *CI*: 1.01-1.22; *P* = 0.039) and $\epsilon 4/\epsilon 4$ genotype had a 1.71-fold risk of developing T2DM (*OR* = 1.71; 95% *CI*: 1.33-2.19; *P* < 0.001) when they were compared with $\epsilon 3/\epsilon 3$ genotype.

Conclusions There is an association between *ApoE* polymorphism and T2DM: allele $\epsilon 4$ and genotypes ($\epsilon 2/\epsilon 2$, $\epsilon 3/\epsilon 4$, and $\epsilon 4/\epsilon 4$) are associated with the increased risk for the development of T2DM, and they may be risk factors for T2DM.

Key words: Apolipoprotein E; Polymorphism; Type 2 diabetes; Meta-analysis

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Biographical note of the first author: CHEN Da Wei, male, born in 1962, PhD, majoring in effects of environmental exposure on health.

INTRODUCTION

It is estimated that only half of the 79 million adults with type 2 diabetes will have adequate access to insulin by 2030 if the current levels of access is not improved^[1]. Moreover, one of the significant causes of worldwide mortality and morbidity is diabetes^[2], especially type 2 diabetes mellitus (T2DM), which is also the major cause of substantial global economic burden^[3]. Therefore, there is an urgent need to identify the important risk factors for T2DM and develop effective strategies to address the problem of T2DM.

It is well accepted that genetic factor, environmental factors, and lifestyle contribute to the development of T2DM. Complex interactions between multiple genes and a range of environmental factors are involved in the onset and progression of type 2 diabetes^[4]. A better understanding of the contribution of genetic factors in the etiology of T2DM will facilitate the development of effective preventive strategies to reduce the ever increasing incidence of T2DM^[5], it will also improve the effectiveness and precision of treatment and prevention strategies^[6].

It is reported that ApoE alleles are important genetic markers for dyslipidaemias^[7], and previous studies indicate that ApoE is among the candidate genes which are most likely associated with CAD in T2DM patients^[8]. *ApoE* draws much attention due to some reports supporting the association between *ApoE* polymorphism and T2DM^[9-11]. In humans, *ApoE* gene is located on the chromosome at position 19q13.2 with 3 isoforms, ApoE2, ApoE3, and ApoE4; and 6 genotypes having 3 homozygous: $\epsilon 2/\epsilon 2$, $\epsilon 3/\epsilon 3$, and $\epsilon 4/\epsilon 4$, and 3 heterozygous: $\epsilon 2/\epsilon 3$, $\epsilon 2/\epsilon 4$, and $\epsilon 3/\epsilon 4$ ^[12]. Besides T2DM, *ApoE* is also involved in many diseases, such as coronary heart disease (CHD)^[13], ischemic cerebrovascular disease (ICD)^[14], and Alzheimer's disease^[15].

Much of the recent research has studied the association between the *ApoE* gene polymorphism and the risk of T2DM, however, there are inconsistencies between the results of the different studies. The inconsistency may result from the difference of included population, sample size, and genotyping methods. Moreover, 18 new papers^[9,16-32] have been published since the publication of latest meta-analysis of the association between *ApoE* gene polymorphism and T2DM in 2014^[33]. Thus, we conducted a further meta-analysis to explore whether *ApoE* polymorphism is associated with the increased risk of T2DM by including these new

published articles.

METHODS

Search Strategy

We performed this meta-analysis by extensive literature search in PubMed, Web of Science, Medline, WanFang, VIP, and CNKI databases (last search on February 28, 2019). We used the following terms for our search strategy, ('ApoE' OR 'Apolipoprotein E') AND ('polymorphism, Genetic' OR 'variant' OR 'mutation') AND ('type 2 diabetes mellitus' OR 'type 2 diabetes' OR 'T2DM' OR 'non-insulin dependent diabetes' OR 'NIDDM'). The equivalent Chinese terms were used in the Chinese databases. In addition, we retrieved related articles that had not been identified in the initial search to replenish literatures.

Inclusion/Exclusion Criteria

Studies included in this meta-analysis were based on the following criteria: (1) case-control studies; (2) assessing the association between *ApoE* polymorphism and type 2 diabetes. The exclusion criteria met the follows: (1) duplicate articles; (2) no healthy controls; (3) insufficient information on genotype or allele frequencies.

Data Extraction

We extracted the main characteristics of each eligible study, including first author's last name, date of publication, region, population's ethnicity, genotyping method, number of cases and controls, and counts of the *ApoE* genotype or allele. Hardy-Weinberg equilibrium (HWE) was collected and calculated among the controls.

Quality Assessment

We used the Newcastle-Ottawa scale (NOS) to assess the quality of each article by a 'star' rating system covering selection, comparability, and exposure. A score of 1 point was awarded for each condition a study met, and no point (0 score) if the condition or requirement was not met. We calculated the total Quality Score of each study. Two authors (Jikang Shi and Shuping Ren) assessed the quality of included studies independently. When inconformity occurred between the two authors, we discussed with the third investigator (CHEN Da Wei) and came to a conformity. We included those studies with poor quality score to avoid selection bias.

Statistical Analysis

We calculated the allele and genotype frequencies of *ApoE* for each study to evaluate the HWE through Goodness of fit Chi-square test among control groups, and $P < 0.05$ was seen as a significant deviation from HWE. The strength of association between *ApoE* polymorphisms and type 2 diabetes susceptibility was assessed using odds ratios (*OR*) and 95% confidence intervals (95% *CI*) because outcome variable was binary. Heterogeneity was assessed by the Chi-square test based Q-statistic and quantified by I^2 -statistic^[34]. Random-effect models (DerSimonian and Laird methods) were applied to calculate *OR* and 95% *CI* when P value of Q test was more than 0.10 or I^2 value was more than 50%; otherwise, fixed-effect models (Mantel and Haenszel methods) were used ($I^2 \geq 50\%$ considered heterogeneity existed in between-study in this meta-analysis). Subgroup analyses stratified by ethnicity, quality score and Hardy–Weinberg equilibrium were calculated to trace main sources of heterogeneity and to identify the association between *ApoE* polymorphisms and type 2 diabetes in different groups. Publication bias was evaluated using funnel plots, and quantified by the Begg's and Egger's tests ($P < 0.05$ considered statistically significant

publication bias)^[35]. Sensitivity analysis was performed to examine stability of results by omitting each study in each turn. All data management and statistical analyses were used R soft-ware (version 3.4.3), P -value < 0.05 was considered statistically significant.

RESULTS

Study Characteristics

Our meta-analysis initially collected 791 published articles, including 782 papers collected by our search strategy and 9 papers through the references. After scanning the abstracts and full texts according to the inclusion and exclusion criteria, we included 59 eligible articles with 6,872 cases and 8,250 controls in this paper. The protocol of the process for literature identification and selection is listed in Figure 1, and the baseline characteristics of the included studies are summarized in Table 1, all the results of meta-analysis is shown in Table 2.

Association between Alleles of *ApoE* and Type 2 Diabetes

We found a significant heterogeneity when we comparing *ApoE* $\epsilon 2$ with $\epsilon 3$ allele ($I^2 = 62\%$), and had

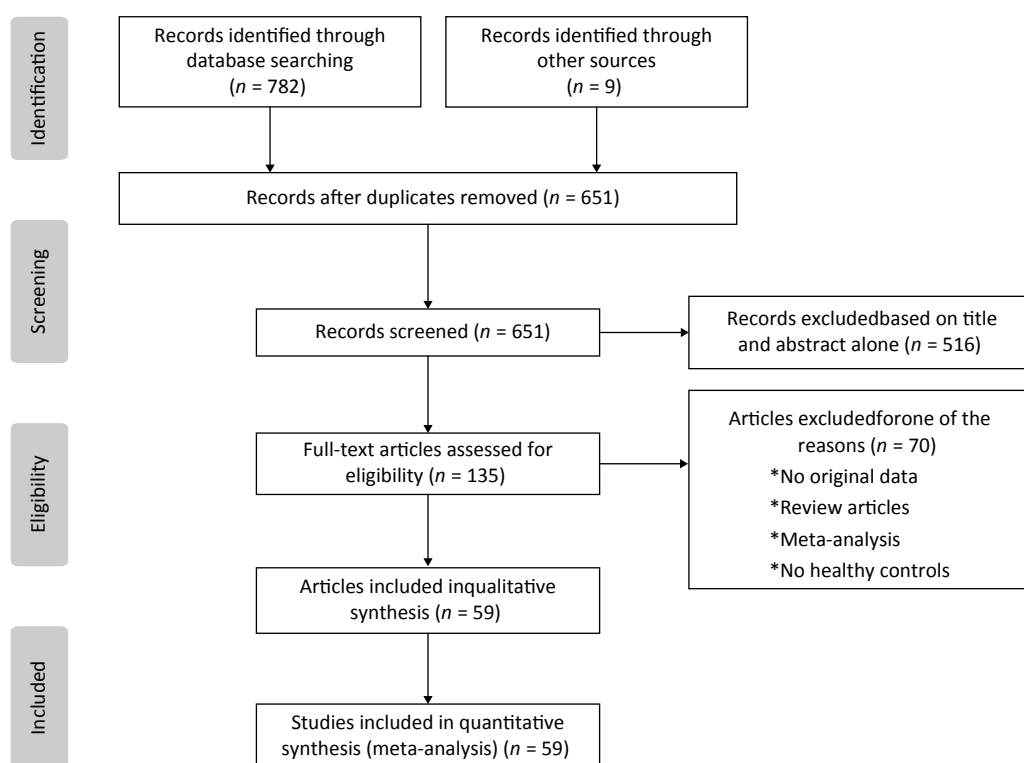


Figure 1. Flow chart of the process for literature identification and selection.

Table 1. Main characteristics of the included studies

| Study | Year | Region | Ethnicity | Genotyping method | Sample size (case/control) | Quality score | HWE Y/N(P) | $\epsilon 2/\epsilon 2(n)+\epsilon 2/\epsilon 3(n)$ | | $\epsilon 2/\epsilon 4(n)+\epsilon 3/\epsilon 3(n)$ | | $\epsilon 3/\epsilon 4(n)+\epsilon 4/\epsilon 4(n)$ | |
|--------------------------------|------|---------|-----------|-------------------------------|----------------------------|---------------|-------------|---|---------|---|---------|---|---------|
| | | | | | | | | case | control | case | control | case | control |
| Singh ^[36] | 2006 | India | Asian | PCR-RELP | 90/97 | 9 | Y(0.184) | 1+4 | 1+7 | 2+78 | 0+74 | 5+0 | 13+2 |
| Al-Majed ^[16] | 2011 | Kuwait | Other | PCR-RELP | 105/62 | 6 | N(0.006) | 7+2 | 2+3 | 2+73 | 2+46 | 6+15 | 9+1 |
| Chaudhary ^[9] | 2012 | Bangkok | Other | PCR-RELP | 155/149 | 8 | Y(0.121) | 1+2 | 2+12 | 1+117 | 0+113 | 30+4 | 21+1 |
| Errera ^[37] | 2006 | Brazil | Other | PCR-RELP | 95/107 | 7 | Y(0.584) | 0+13 | 0+7 | 2+68 | 0+77 | 12+0 | 23+0 |
| Alharbi ^[17] | 2014 | Riyadh | Other | TaqMan | 438/460 | 7 | N(< 0.001) | 35+26 | 27+18 | 13+290 | 11+334 | 35+39 | 60+10 |
| Inamdar ^[38] | 2000 | India | Asian | Flat gel isoelectric focusing | 60/40 | 8 | Y(0.054) | 2+8 | 1+9 | 3+17 | 2+10 | 16+14 | 8+10 |
| Kwon ^[39] | 2007 | Korea | Asian | PCR-RELP | 94/88 | 7 | Y(0.924) | 0+13 | 0+5 | 3+63 | 0+70 | 14+1 | 12+1 |
| Atta ^[18] | 2016 | Egypt | Other | PCR-RELP | 45/45 | 5 | Y(0.098) | 0+12 | 0+3 | 12+12 | 3+30 | 9+0 | 9+0 |
| Vauhkonen ^[40] | 1997 | Finland | Caucasian | PCR-RELP | 86/125 | 8 | Y(0.963) | 0+7 | 0+9 | 3+48 | 2+76 | 20+8 | 33+5 |
| Erdogan ^[19] | 2009 | Turkey | Caucasian | PCR-RELP | 56/35 | 7 | N(< 0.001) | 0+4 | 0+0 | 0+40 | 0+28 | 12+0 | 7+0 |
| Eto ^[41] | 1986 | Japan | Asian | Flat gel isoelectric focusing | 105/111 | 8 | Y(0.339) | 0+9 | 1+10 | 0+73 | 1+80 | 21+2 | 16+3 |
| Guan ^[42] | 2009 | China | Asian | PCR-LDR | 213/111 | 7 | Y(0.499) | 8+32 | 1+32 | 7+141 | 1+88 | 24+1 | 9+1 |
| Leiva ^[43] | 2005 | Chile | Other | PCR-RELP | 193/139 | 7 | Y(0.293) | 0+12 | 0+10 | 4+133 | 3+87 | 43+1 | 39+0 |
| Liu ^[44] | 2003 | China | Asian | PCR-RELP | 80/81 | 7 | Y(0.217) | 0+11 | 0+4 | 1+56 | 2+64 | 12+0 | 11+0 |
| Mehmet ^[20] | 2015 | Turkey | Caucasian | PCR-RELP | 100/50 | 8 | N(0.039) | 0+6 | 0+22 | 0+81 | 0+19 | 13+0 | 9+0 |
| Xie ^[45] | 2011 | China | Asian | PCR-RELP | 60/20 | 7 | Y(0.936) | 0+13 | 1+3 | 4+8 | 2+8 | 19+16 | 5+1 |
| Mustapic ^[21] | 2012 | Croatia | Caucasian | TaqMan | 196/456 | 6 | Y(0.331) | 0+35 | 1+48 | 2+127 | 2+328 | 30+2 | 76+1 |
| Santos ^[46] | 2002 | Mexico | Other | PCR-RELP | 36/22 | 8 | Y(0.423) | 0+0 | 1+2 | 0+32 | 1+10 | 3+1 | 8+0 |
| Kamboh ^[47] | 1995 | USA | Caucasian | IEF-immunoblottin and PCR | 116/659 | 6 | Y(0.992) | 0+23 | 6+88 | 5+62 | 19+382 | 26+0 | 150+14 |
| Ng ^[48] | 2016 | China | Asian | Other | 386/200 | 6 | Y(0.168) | 4+53 | 1+32 | 5+282 | 6+142 | 39+3 | 19+0 |
| Eto ^[49] | 1995 | Japan | Asian | Flat gel isoelectric focusing | 281/576 | 8 | Y(0.609) | 1+25 | 2+35 | 1+192 | 4+414 | 55+7 | 111+10 |
| Morbois Trabut ^[50] | 2006 | France | Caucasian | PCR-RELP | 210/481 | 7 | Y(0.773) | 2+31 | 5+71 | 1+143 | 14+294 | 33+0 | 87+10 |
| Powell ^[51] | 2003 | UK | Caucasian | PCR-RELP | 187/102 | 7 | Y(0.094) | 3+22 | 2+7 | 3+89 | 1+57 | 27+3 | 21+0 |
| Guangda ^[52] | 1999 | China | Asian | PCR-RELP | 89/72 | 7 | Y(0.122) | 1+13 | 1+7 | 1+66 | 2+53 | 7+1 | 7+2 |
| Zhang ^[53] | 2000 | China | Asian | PCR-RELP | 63/71 | 8 | N(0.009) | 0+7 | 0+5 | 0+50 | 3+56 | 6+0 | 6+0 |
| Zhang ^[54] | 2003 | China | Asian | PCR-RELP | 74/191 | 8 | Y(0.878) | 0+5 | 1+23 | 1+55 | 1+134 | 12+1 | 31+1 |
| Sun ^[55] | 2013 | China | Asian | PCR-RELP | 243/78 | 7 | Y(0.414) | 6+36 | 2+12 | 0+180 | 1+55 | 21+0 | 6+1 |
| Hua ^[56] | 2006 | China | Asian | PCR-RELP | 50/60 | 8 | Y(0.190) | 2+4 | 0+7 | 4+68 | 2+75 | 20+2 | 13+3 |
| Guo ^[57] | 2003 | China | Asian | PCR-RELP | 40/52 | 7 | Y(0.739) | 0+4 | 0+5 | 2+23 | 1+39 | 9+2 | 6+1 |
| Liang ^[23] | 2017 | China | Asian | PCR-RELP | 44/374 | 6 | Y(0.816) | 1+3 | 5+57 | 1+31 | 6+267 | 7+1 | 38+1 |
| Shen ^[58] | 2002 | China | Asian | PCR-RELP | 106/110 | 7 | Y(0.577) | 1+7 | 1+12 | 2+84 | 4+74 | 11+1 | 18+1 |
| Zheng ^[59] | 1998 | China | Asian | PCR-RELP | 112/60 | 8 | Y(0.801) | 2+16 | 1+8 | 1+81 | 0+45 | 11+1 | 6+0 |
| Hua ^[60] | 2004 | China | Asian | PCR-RELP | 38/60 | 7 | Y(0.434) | 1+7 | 0+4 | 2+24 | 1+45 | 4+0 | 8+2 |
| Liu ^[24] | 2014 | China | Asian | PCR-RELP | 215/298 | 7 | N(< 0.001)) | 10+0 | 2+0 | 0+174 | 0+272 | 31+0 | 23+1 |

| Study | Year | Region | Ethnicity | Genotyping method | Sample size (case/control) | Quality score | HWE Y/N(P) | Continued | | | | | |
|-----------------------|------|--------|-----------|-------------------|----------------------------|---------------|------------|-----------------------------|------|-----------------------------|-------|-----------------------------|------|
| | | | | | | | | $\epsilon 2/\epsilon 2(n)+$ | | $\epsilon 2/\epsilon 4(n)+$ | | $\epsilon 3/\epsilon 4(n)+$ | |
| | | | | | | | | $\epsilon 2/\epsilon 3(n)$ | | $\epsilon 3/\epsilon 3(n)$ | | $\epsilon 4/\epsilon 4(n)$ | |
| Xiang ^[61] | 1995 | China | Asian | PCR-RELP | 125/50 | 7 | Y(0.715) | 2+16 | 0+4 | 0+78 | 1+38 | 26+3 | 6+1 |
| Chen ^[25] | 2006 | China | Asian | PCR-RELP | 97/105 | 7 | Y(0.906) | 2+15 | 1+18 | 1+70 | 2+72 | 8+1 | 10+1 |
| Xiang ^[62] | 1999 | China | Asian | PCR-ASO | 130/50 | 8 | Y(0.715) | 3+14 | 0+4 | 1+85 | 1+38 | 24+3 | 6+1 |
| Shen ^[63] | 2002 | China | Asian | PCR-RELP | 35/50 | 6 | Y(0.112) | 3+11 | 0+6 | 2+4 | 4+31 | 14+0 | 9+0 |
| Xiong ^[26] | 2013 | China | Asian | PCR-RELP | 121/112 | 8 | Y(0.991) | 0+15 | 1+13 | 1+72 | 2+72 | 31+2 | 22+2 |
| Zhou ^[64] | 2005 | China | Asian | PCR-RELP | 67/68 | 7 | Y(0.263) | 0+13 | 2+9 | 1+47 | 0+46 | 6+0 | 11+0 |
| Xiang ^[65] | 2005 | China | Asian | PCR-ASO | 101/95 | 7 | Y(0.438) | 1+10 | 1+10 | 1+65 | 1+65 | 20+4 | 15+3 |
| Long ^[66] | 1999 | China | Asian | PCR-RELP | 67/135 | 7 | Y(0.124) | 0+15 | 0+18 | 3+36 | 4+101 | 12+1 | 12+0 |
| Liang ^[67] | 2005 | China | Asian | PCR-RELP | 145/90 | 8 | Y(0.592) | 0+17 | 0+12 | 6+102 | 2+68 | 18+2 | 8+0 |
| Gu ^[68] | 2004 | China | Asian | PCR-RELP | 63/90 | 8 | Y(0.592) | 0+9 | 0+12 | 3+43 | 2+68 | 7+1 | 8+0 |
| Yang ^[69] | 1995 | China | Asian | PCR-RELP | 125/50 | 7 | N(0.028) | 2+16 | 1+3 | 0+78 | 1+38 | 26+3 | 5+2 |
| Rong ^[32] | 2013 | China | Asian | PCR-RELP | 18/29 | 7 | Y(0.953) | 0+4 | 0+8 | 0+18 | 0+29 | 2+0 | 1+0 |
| Liu ^[27] | 2016 | China | Asian | PCR-RELP | 300/300 | 8 | N(< 0.001) | 14+0 | 2+0 | 0+243 | 0+274 | 43+0 | 23+1 |
| Tang ^[28] | 2007 | China | Asian | PCR-RELP | 41/60 | 6 | Y(0.80) | 0+1 | 0+3 | 2+28 | 1+43 | 10+0 | 13+0 |
| Qiu ^[70] | 2008 | China | Asian | PCR-RELP | 129/110 | 8 | Y(0.481) | 0+14 | 1+18 | 3+95 | 2+76 | 14+3 | 11+2 |
| Guo ^[71] | 2007 | China | Asian | ARMS-PCR | 40/40 | 6 | Y(0.618) | 0+1 | 1+4 | 3+29 | 1+27 | 7+1 | 7+0 |
| Xiong ^[72] | 2008 | China | Asian | MultiARMS PCR | 316/512 | 6 | Y(0.744) | 2+18 | 3+48 | 6+230 | 9+359 | 47+13 | 87+6 |
| Ge ^[29] | 2013 | China | Asian | PCR-RELP | 200/210 | 7 | Y(0.544) | 3+35 | 8+40 | 2+86 | 8+103 | 73+1 | 47+4 |
| Xiang ^[73] | 2010 | China | Asian | PCR-RELP | 41/102 | 7 | Y(0.473) | 0+5 | 0+13 | 1+28 | 0+70 | 7+0 | 19+0 |
| Luo ^[30] | 2016 | China | Asian | PCR-RELP | 35/50 | 6 | N(0.005) | 0+3 | 0+2 | 1+28 | 3+38 | 2+1 | 7+0 |
| Zhang ^[74] | 2007 | China | Asian | PCR-RELP | 38/49 | 6 | N(0.015) | 0+2 | 0+1 | 0+32 | 2+39 | 3+1 | 7+0 |
| Wang ^[31] | 2014 | China | Asian | PCR-RELP | 57/55 | 8 | N(0.027) | 0+4 | 2+7 | 2+33 | 4+28 | 13+5 | 8+6 |
| Zhang ^[75] | 1999 | China | Asian | PCR-RELP | 56/76 | 5 | Y(0.631) | 0+3 | 1+7 | 1+40 | 2+55 | 11+1 | 11+1 |
| Xiong ^[76] | 2005 | China | Asian | PCR-RELP | 32/30 | 7 | Y(0.608) | 1+5 | 0+4 | 1+22 | 1+23 | 2+1 | 2+0 |
| Dai ^[77] | 2000 | China | Asian | PCR-RELP | 32/90 | 8 | Y(0.253) | 0+5 | 0+14 | 0+23 | 1+64 | 3+1 | 9+2 |

Note. HWE, Hardy-Weinberg equilibrium.

Table 2. Meta-analysis results of association between ApoE polymorphism and type 2 diabetes

| Variable | OR (95% CI) | I ² (%) | P |
|-------------------------|-------------------|--------------------|---------|
| <i>ApoE</i> alleles | | | |
| $\epsilon 2$ | 1.16 (0.98, 1.37) | 62 | 0.079 |
| $\epsilon 4$ | 1.18 (1.09, 1.28) | 36 | < 0.001 |
| <i>ApoE</i> genotypes | | | |
| $\epsilon 2/\epsilon 2$ | 1.46 (1.11, 1.93) | 0 | 0.007 |
| $\epsilon 2/\epsilon 3$ | 1.09 (0.90, 1.32) | 55 | 0.397 |
| $\epsilon 2/\epsilon 4$ | 1.15 (0.90, 1.46) | 0 | 0.276 |
| $\epsilon 3/\epsilon 4$ | 1.11 (1.01, 1.22) | 39 | 0.039 |
| $\epsilon 4/\epsilon 4$ | 1.71 (1.33, 2.19) | 0 | < 0.001 |

Note. ApoE alleles ($\epsilon 2$ and $\epsilon 4$) and genotypes ($\epsilon 2/\epsilon 2$, $\epsilon 2/\epsilon 3$, $\epsilon 2/\epsilon 4$, $\epsilon 3/\epsilon 4$, and $\epsilon 4/\epsilon 4$) were compared with $\epsilon 3$ and $\epsilon 3/\epsilon 3$.

the pooled OR of 1.16 (95% CI: 0.98-1.37; $P = 0.079$) calculated by the random-effects model (Figure 2); however, there was not heterogeneity in the comparison of *ApoE* $\epsilon 4$ with $\epsilon 3$ allele ($I^2 = 36\%$), and the pooled OR was 1.18 (95% CI: 1.09-1.28; $P < 0.001$) when the fixed-effects model was applied to compare *ApoE* $\epsilon 4$ with $\epsilon 3$ (Figure 3), indicating that *ApoE* $\epsilon 4$ allele may be a risk factor for type 2 diabetes.

Association between Genotypes of *ApoE* and Type 2 Diabetes

There were five genotypes ($\epsilon 2/\epsilon 2$, $\epsilon 2/\epsilon 3$, $\epsilon 2/\epsilon 4$, $\epsilon 3/\epsilon 4$, and $\epsilon 4/\epsilon 4$) were compared with $\epsilon 3/\epsilon 3$ genotype. No significant heterogeneity was found when the comparison was performed between the $\epsilon 2/\epsilon 2$ and $\epsilon 3/\epsilon 3$ genotypes ($I^2 = 0\%$), and the yielded OR of $\epsilon 2/\epsilon 2$ genotype versus $\epsilon 3/\epsilon 3$ genotype using a fixed-effects model was 1.46 (95% CI: 1.11-1.93; $P = 0.007$) (Figure 4), indicating that the $\epsilon 2/\epsilon 2$ genotype might produce a harmful effect on type 2 diabetes. However, when $\epsilon 2/\epsilon 3$ genotype was compared with $\epsilon 3/\epsilon 3$ genotype, there was significant heterogeneity ($I^2 = 55\%$), and the yielded OR of $\epsilon 2/\epsilon 3$ genotype versus $\epsilon 3/\epsilon 3$ genotype using a random-effects model was 1.09 (95% CI: 0.90-1.32; $P = 0.397$) (Figure 5). Compared with $\epsilon 3/\epsilon 3$ genotype, there were no significant heterogeneity between $\epsilon 2/\epsilon 4$, $\epsilon 3/\epsilon 4$, and $\epsilon 4/\epsilon 4$ genotype, respectively ($I^2 = 0\%$, $I^2 = 39\%$, and $I^2 = 0\%$). The yielded OR of $\epsilon 2/\epsilon 4$ genotype versus $\epsilon 3/\epsilon 3$ genotype using a fixed-effects model was 1.15 (95% CI: 0.90-1.46; $P = 0.276$) (Figure 6). The yielded OR of $\epsilon 3/\epsilon 4$ genotype versus $\epsilon 3/\epsilon 3$ genotype using a fixed-effects model was 1.11 (95% CI: 1.01-1.22; $P = 0.039$) (Figure 7). For the comparison of $\epsilon 4/\epsilon 4$ genotype with $\epsilon 3/\epsilon 3$ genotype, the yielded OR showed a 1.71-fold risk of type 2 diabetes (OR = 1.71; 95% CI: 1.33-2.19; $P < 0.001$) using the fixed-effects model (Figure 8).

Subgroup Analysis

We conducted subgroup analysis stratified by ethnicity, quality score and Hardy–Weinberg equilibrium in order to identify main sources of heterogeneity. There were significant heterogeneity in the comparison of *ApoE* $\epsilon 2$ with $\epsilon 3$ allele ($I^2 = 62\%$) and the comparison of $\epsilon 2/\epsilon 3$ genotype with $\epsilon 3/\epsilon 3$ genotype ($I^2 = 55\%$) in our paper; however, we could not identify the sources of heterogeneity and there was no significant association between *ApoE* polymorphisms and type 2 diabetes in different subgroups (Supplementary Figures S1-S3, available in www.besjournal.com).

Publication Bias

Funnel plots was used to assess and Begg's and Egger's tests to quantify the publication bias. All the funnel plots for *ApoE* allele and *ApoE* genotypes seemed symmetrical (Supplementary Figures S4-S5, available in www.besjournal.com), and the results of Begg's and Egger's tests revealed that no publication bias was present for the association between *ApoE* allele and type 2 diabetes and between the *ApoE* genotypes and type 2 diabetes (all $P > 0.05$).

Sensitivity Analysis

According to our results of sensitivity analysis, no individual study produced influence on the corresponding pooled ORs and 95% CIs in the comparison of *ApoE* allele with $\epsilon 3$ allele or in the comparison of *ApoE* genotypes with genotype $\epsilon 3/\epsilon 3$ genotype, which indicated these results were relatively stable and credible.

DISCUSSION

In this meta-analysis, we included 59 literatures with 6,872 cases and 8,250 controls to explore the association between the *ApoE* gene polymorphism and type 2 diabetes mellitus. The major findings of our study are that allele $\epsilon 4$ and genotypes ($\epsilon 2/\epsilon 2$, $\epsilon 3/\epsilon 4$, and $\epsilon 4/\epsilon 4$) are associated with the increased risk for the development of T2DM, however, allele $\epsilon 2$ and genotypes ($\epsilon 2/\epsilon 3$ and $\epsilon 2/\epsilon 4$) are not associated with T2DM.

The findings of our meta-analysis are in accordance with the previous studies^[33,78-80], showing that both *ApoE* $\epsilon 4$ allele and the genotypes ($\epsilon 3/\epsilon 4$ and $\epsilon 4/\epsilon 4$) were associated with increased risk of T2DM. Subjects carrying the $\epsilon 4$ alleles had higher plasma total cholesterol levels compared to subjects carrying the $\epsilon 3/\epsilon 3$ genotype, and HDL cholesterol was significantly lower in the $\epsilon 3/\epsilon 4$ than in the $\epsilon 3/\epsilon 3$ individuals^[81]; individuals carrying the $\epsilon 2/\epsilon 2$ genotype had about 31% lower mean LDL than those with the $\epsilon 4/\epsilon 4$ genotype^[82]. Insulin resistance is known to be strongly associated with metabolic dyslipidemia and the correlation of lipid profiles with diabetic phenotypes is significant. Therefore, *ApoE* $\epsilon 4$ allele and the genotypes ($\epsilon 3/\epsilon 4$ and $\epsilon 4/\epsilon 4$) were associated with an increased risk of T2DM through affecting the lipid metabolism.

We found the genotype $\epsilon 2/\epsilon 2$ was associated with increased risk of T2DM, but not allele $\epsilon 2$ or genotype $\epsilon 2/\epsilon 3$; which are not in agreement with

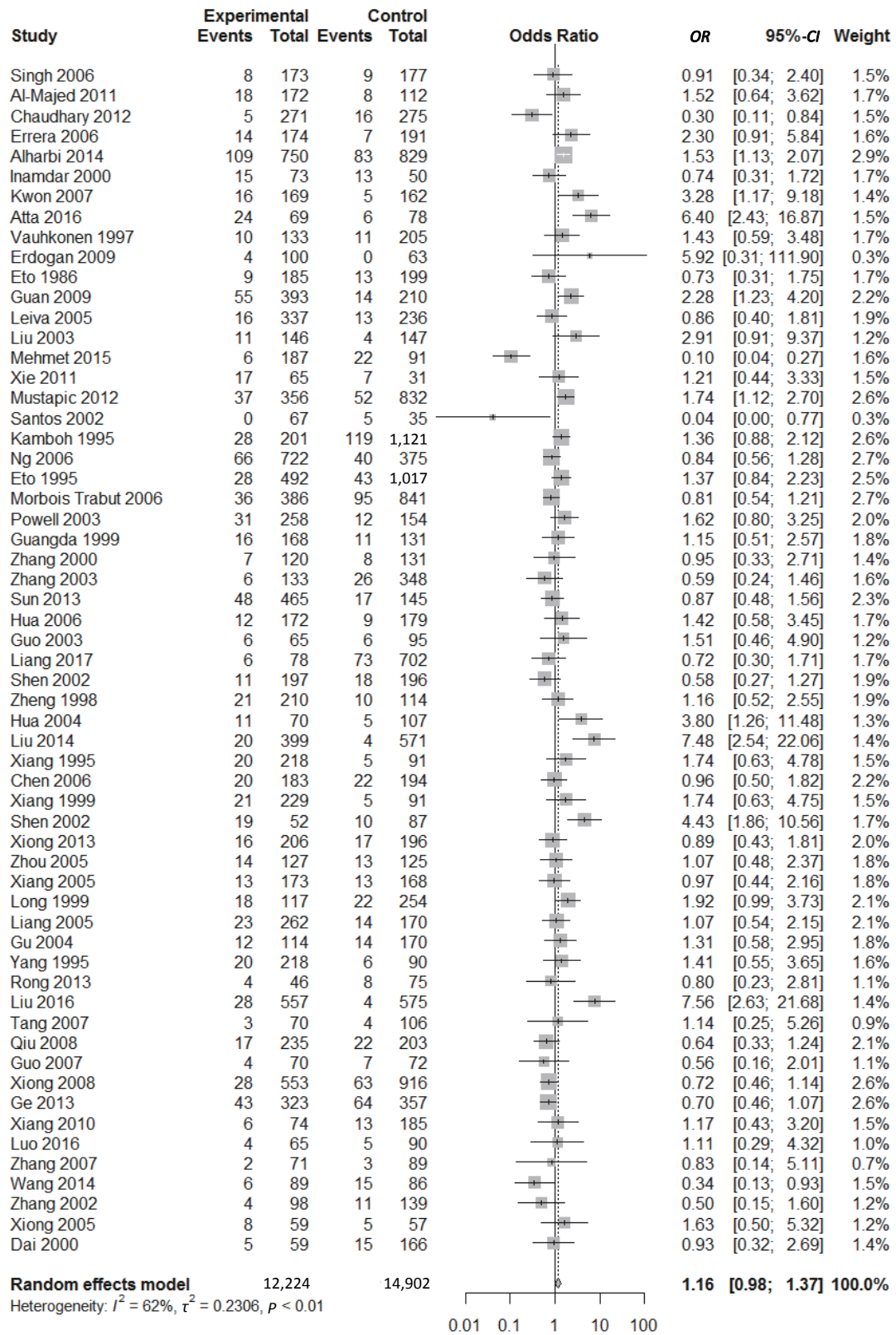


Figure 2. Forest plot for the result of association between type 2 diabetes and ApoE ε2 allele vs. ε3 allele based on a random-effects model.

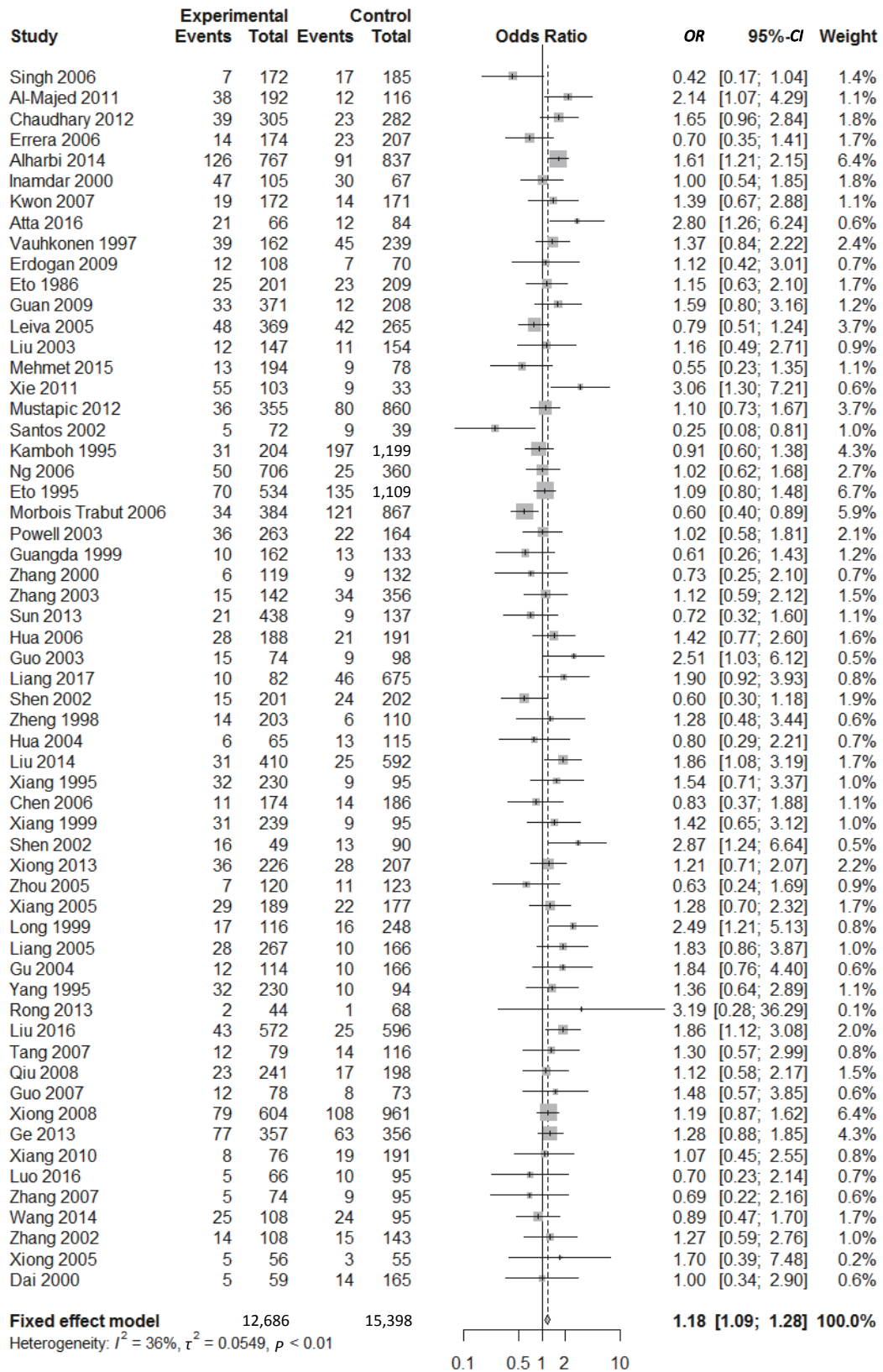


Figure 3. Forest plot for the result of association between type 2 diabetes and *ApoE* $\epsilon 4$ allele vs. $\epsilon 3$ allele based on a fixed-effects model.

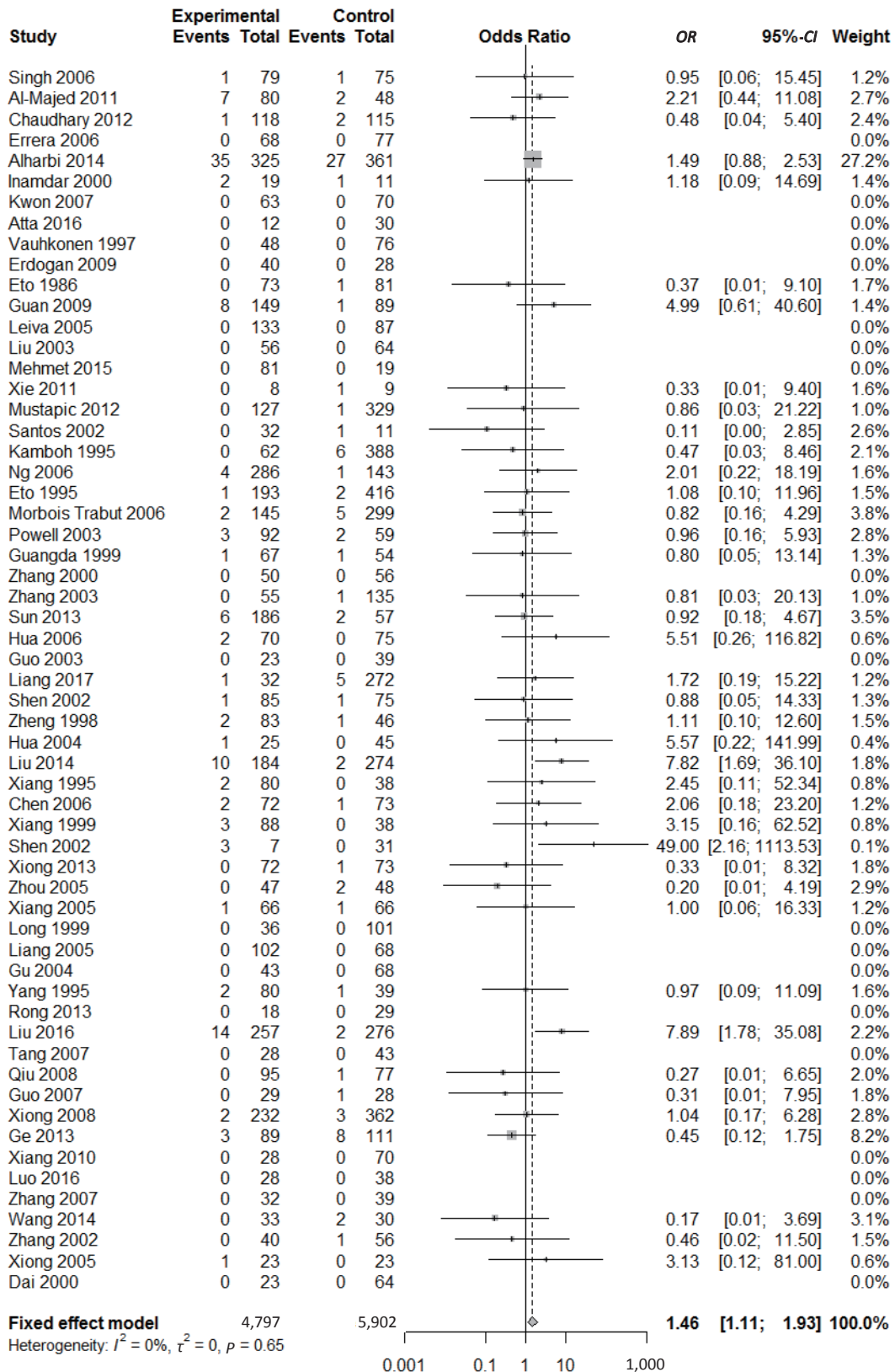


Figure 4. Forest plot for the result of association between type 2 diabetes and ApoE ε2/ε2 genotype vs. ε3/ε3 genotype based on a fixed-effects model.

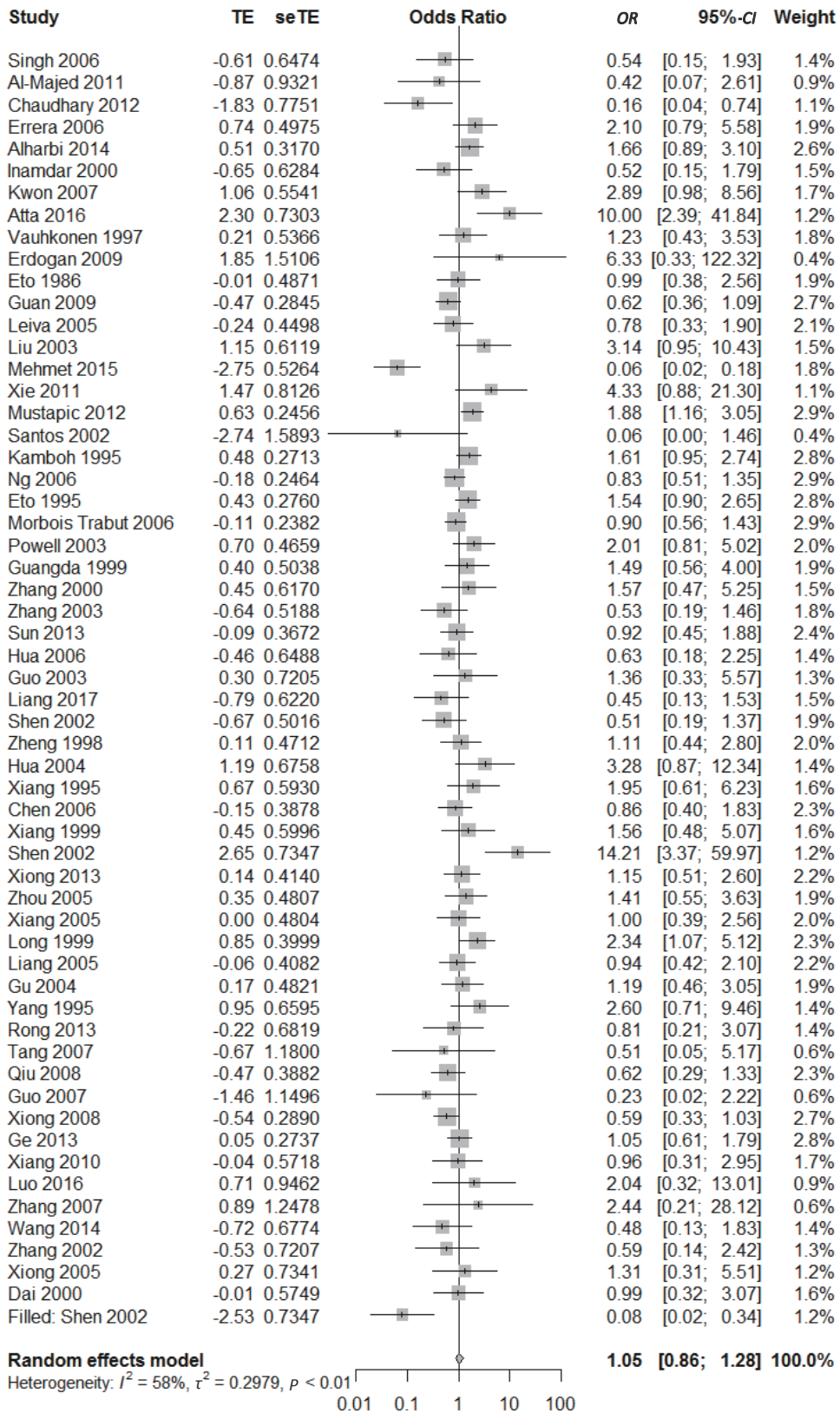


Figure 5. Forest plot for the result of association between type 2 diabetes and ApoE ε2/ε3 genotype vs. ε3/ε3 genotype based on a random-effects model.

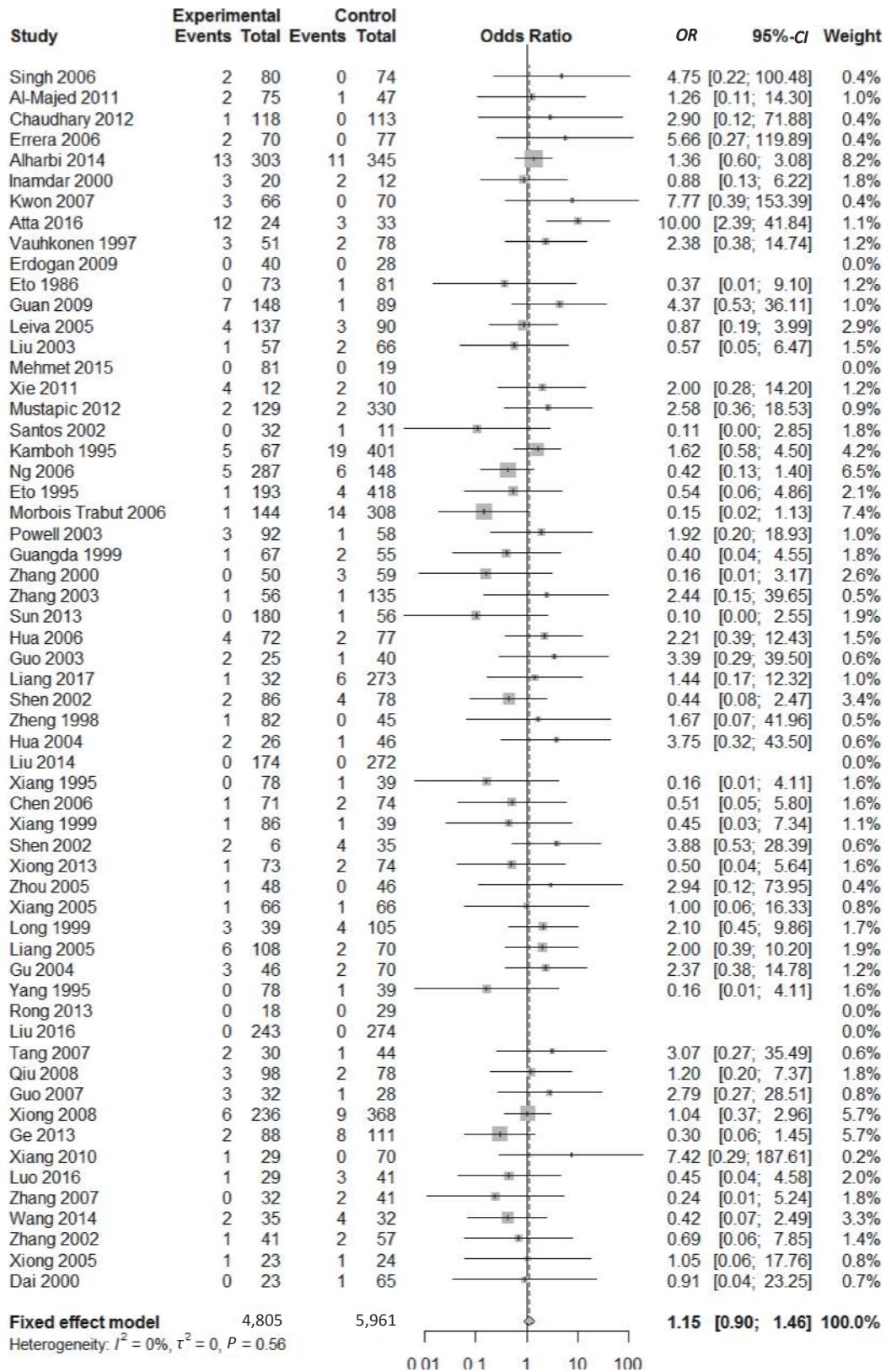


Figure 6. Forest plot for the result of association between type 2 diabetes and ApoE ε2/ε4 genotype vs. ε3/ε3 genotype based on a fixed-effects model.

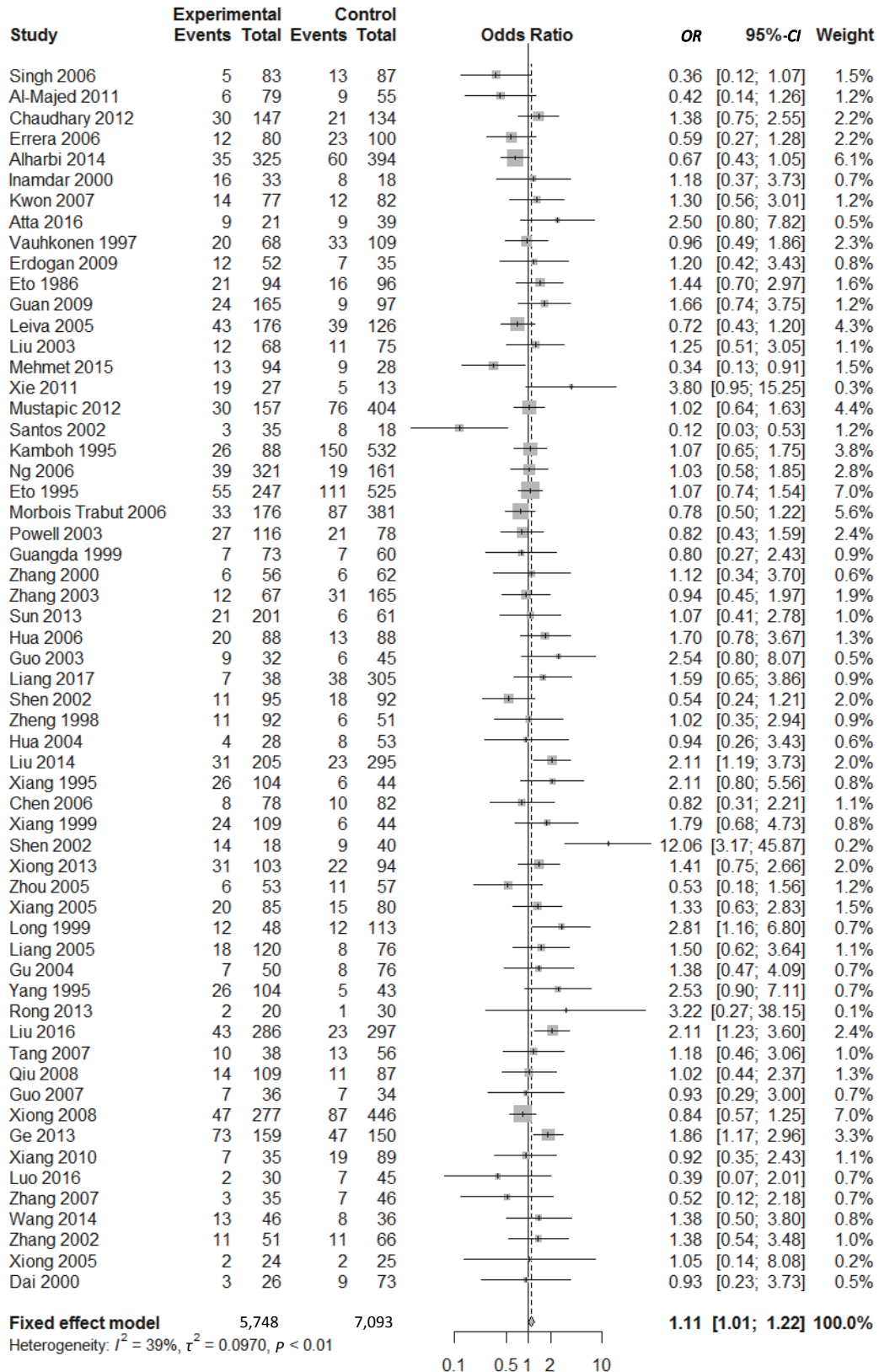


Figure 7. Forest plot for the result of association between type 2 diabetes and ApoE ε3/ε4 genotype vs. ε3/ε3 genotype based on a fixed-effects model.

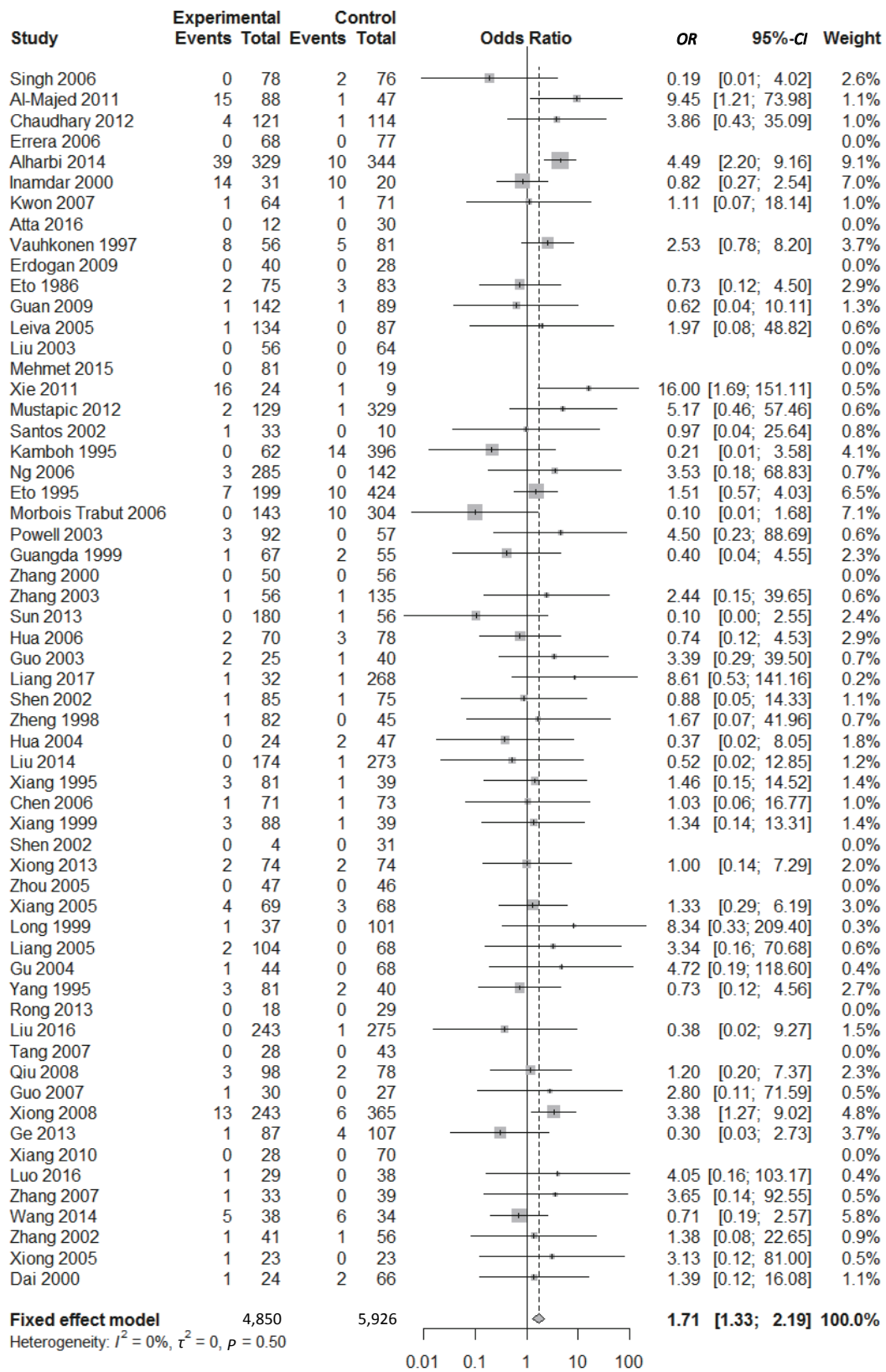


Figure 8. Forest plot for the result of association between type 2 diabetes and ApoE ε4/ε4 genotype vs. ε3/ε3 genotype based on a fixed-effects model.

the results of previous meta-analyses^[33]. The results from Yan et al. showed that $\epsilon 2$ and genotype $\epsilon 2/\epsilon 3$ were associated with increased risk of T2DM, genotype $\epsilon 2/\epsilon 2$ was not associated with increased risk of T2DM. The inconsistency may be caused by the different subjects included. Yan et al. research included only Chinese Han. Furthermore, we did not reveal the difference in the association of ApoE gene polymorphism with T2DM between ethnicities through subgroup analysis. In addition, our findings are consistent with those of Anthopoulos et al. study^[78] which reveals that the ORs for the other $\epsilon 2$ -carriers genotypes ($\epsilon 2/\epsilon 2$, $\epsilon 2/\epsilon 3$, and $\epsilon 2/\epsilon 4$) compared to $\epsilon 3/\epsilon 3$ were greater than 1.00. The slight difference between the present study and Anthopoulos et al' is that the OR of $\epsilon 2/\epsilon 2$ in our study reaches statistical significance while the OR of $\epsilon 2/\epsilon 3$ in Anthopoulos et al' reaches statistical significance. However, the estimates of the results from Anthopoulos et al' study are likely to be attenuated due to the small sample size. Our findings demonstrate that individuals with the genotype carrying single allele $\epsilon 2$ ($\epsilon 2/\epsilon 3$ and $\epsilon 2/\epsilon 4$) are not at the risk of T2DM while those carrying two $\epsilon 2$ allele ($\epsilon 2/\epsilon 2$) possess higher risk for T2DM, which also coincides with the finding that the higher frequency of the $\epsilon 2/APOE$ allele might be primarily related to T2DM^[37].

The strengths of the present study are that, 1) we included all the published literatures on the association between *ApoE* gene polymorphism and T2DM regardless of regions or ethnicities; 2) we had a large sample size. There are 18 new published papers discussing the association between *ApoE* gene polymorphism and T2DM since the last meta-analysis published in 2014, all of them are included in our present meta-analysis, which will provide more convincing evidence to the association of *ApoE* gene polymorphism with T2DM; 3) the results of our sensitivity analysis demonstrate that the conclusion of the present study is very stable; 4) the results of publication bias analysis reveal that the conclusion of our study is absent of publication bias. However, our study also has several weaknesses, 1) presence of heterogeneity in our study. We did the subgroup analysis on HWE, genotyping methods and ethnicities, but we did not trace the source of heterogeneity; 2) since the present study is a case-control study, the findings of our study cannot provide the causal relationship between *ApoE* gene polymorphism and T2DM, only the association of *ApoE* gene polymorphism with T2DM.

CONCLUSION

There is an association between *ApoE* polymorphism and T2DM: allele $\epsilon 4$ and genotypes ($\epsilon 2/\epsilon 2$, $\epsilon 3/\epsilon 4$, and $\epsilon 4/\epsilon 4$) are associated with the increased risk for the development of T2DM, and they may be risk factors for T2DM.

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AUTHOR CONTRIBUTIONS

REN Shu Ping conceived and designed the study; CHEN Da Wei, SHI Ji Kang, LI Yun, and YANG Yu collected, and assembled the data; CHEN Da Wei and SHI Ji Kang analyzed and interpreted the data; CHEN Da Wei, SHI Ji Kang, and REN Shu Ping contributed to the writing process.

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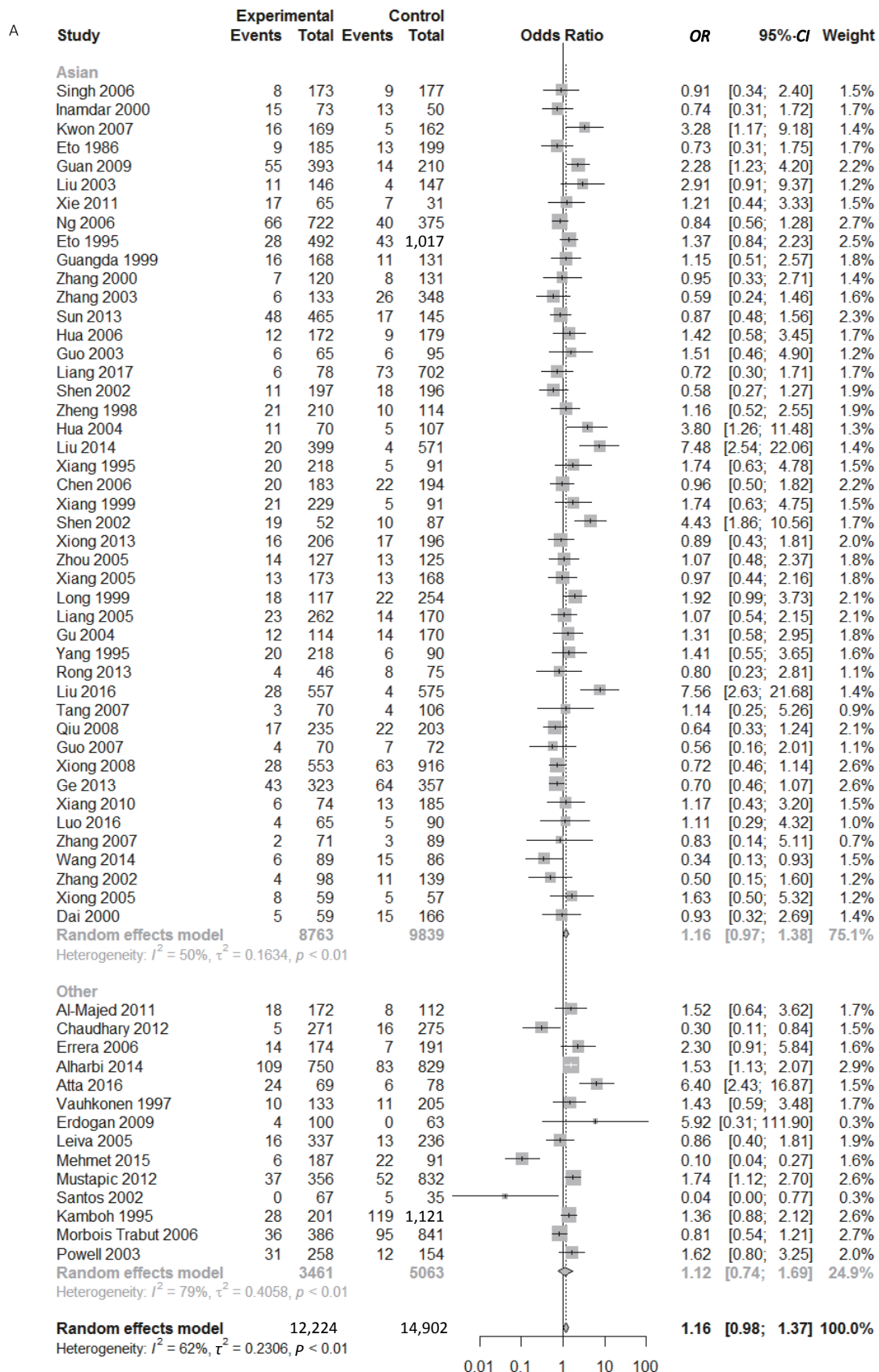
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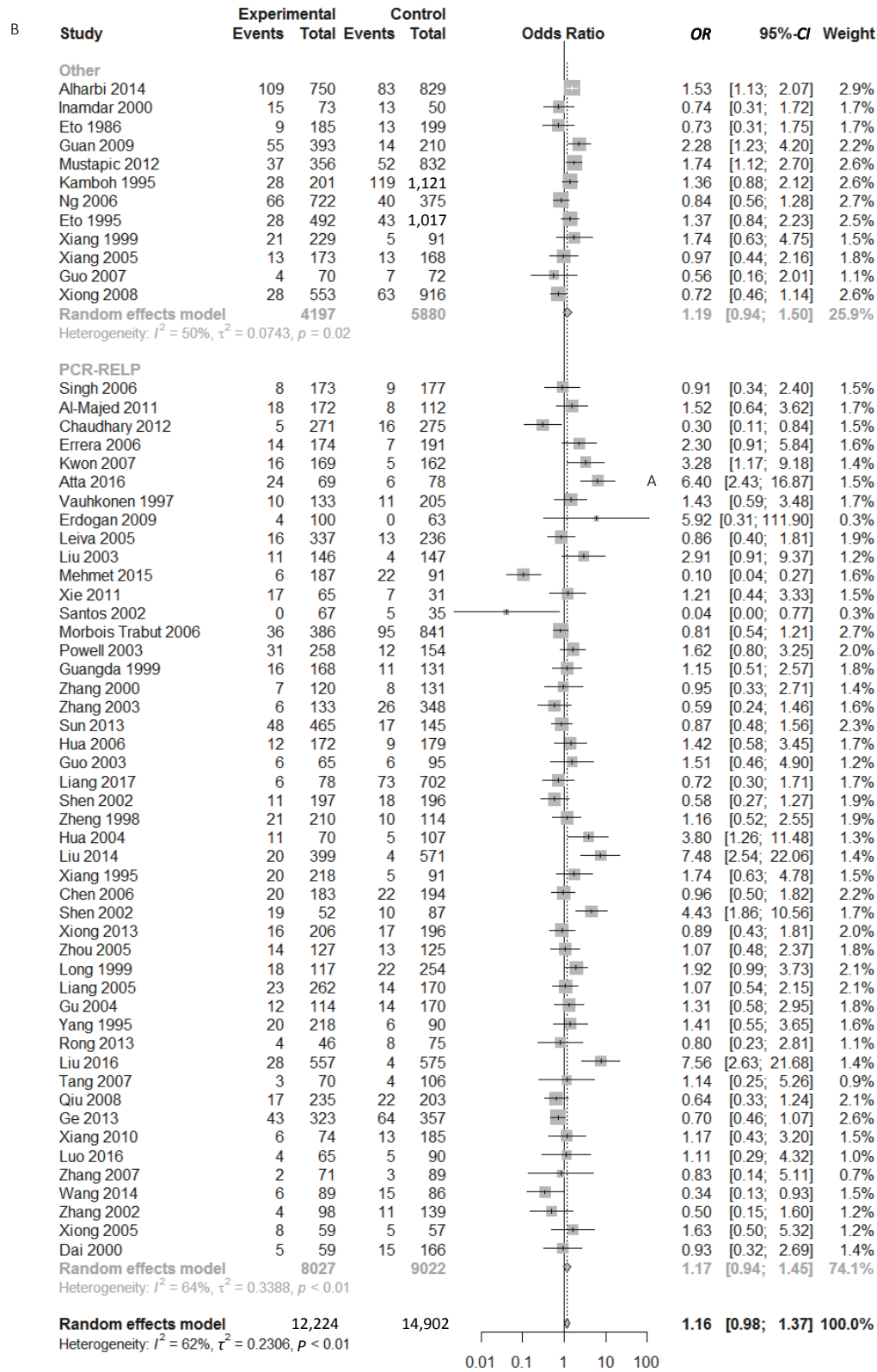
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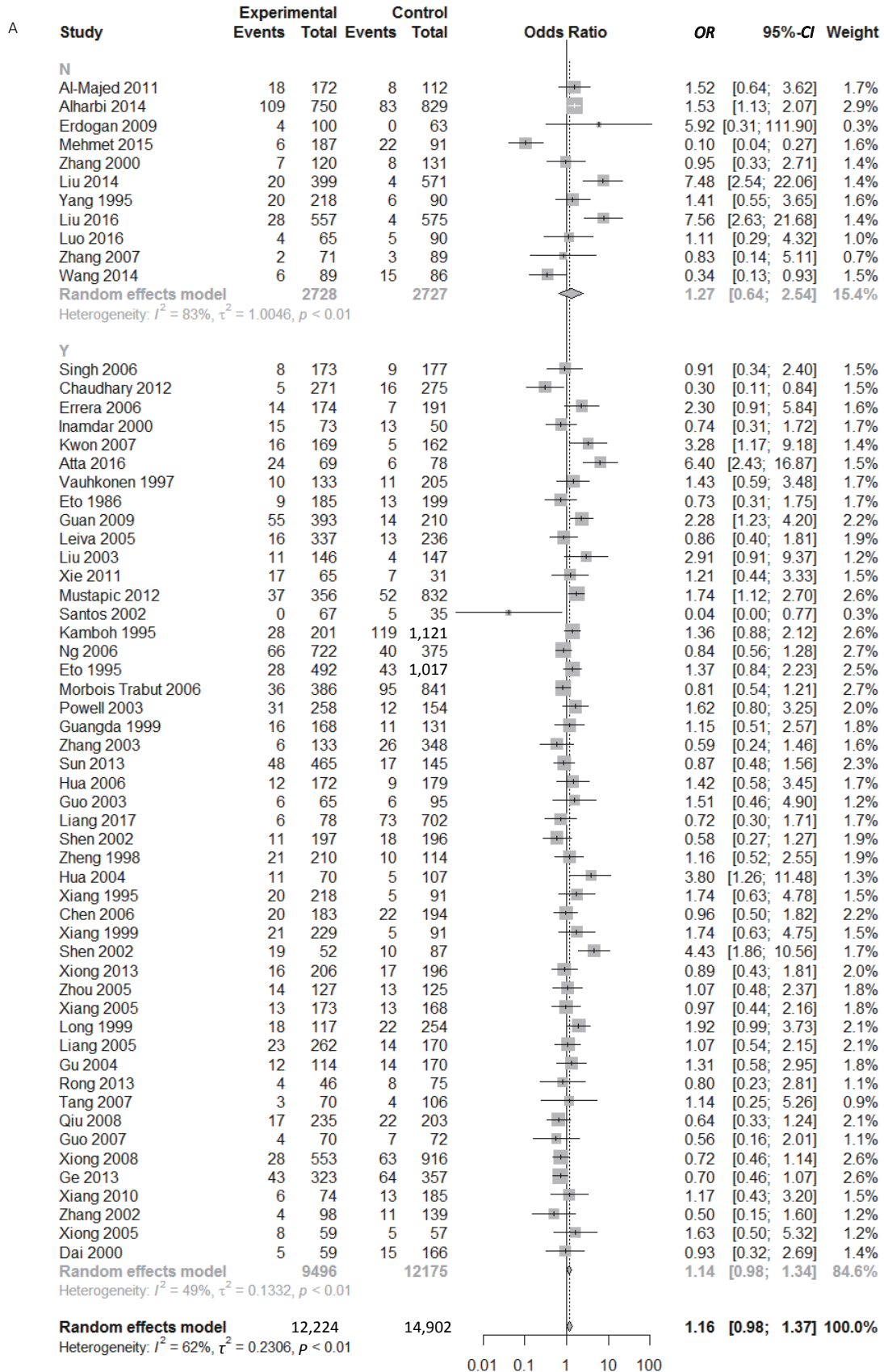
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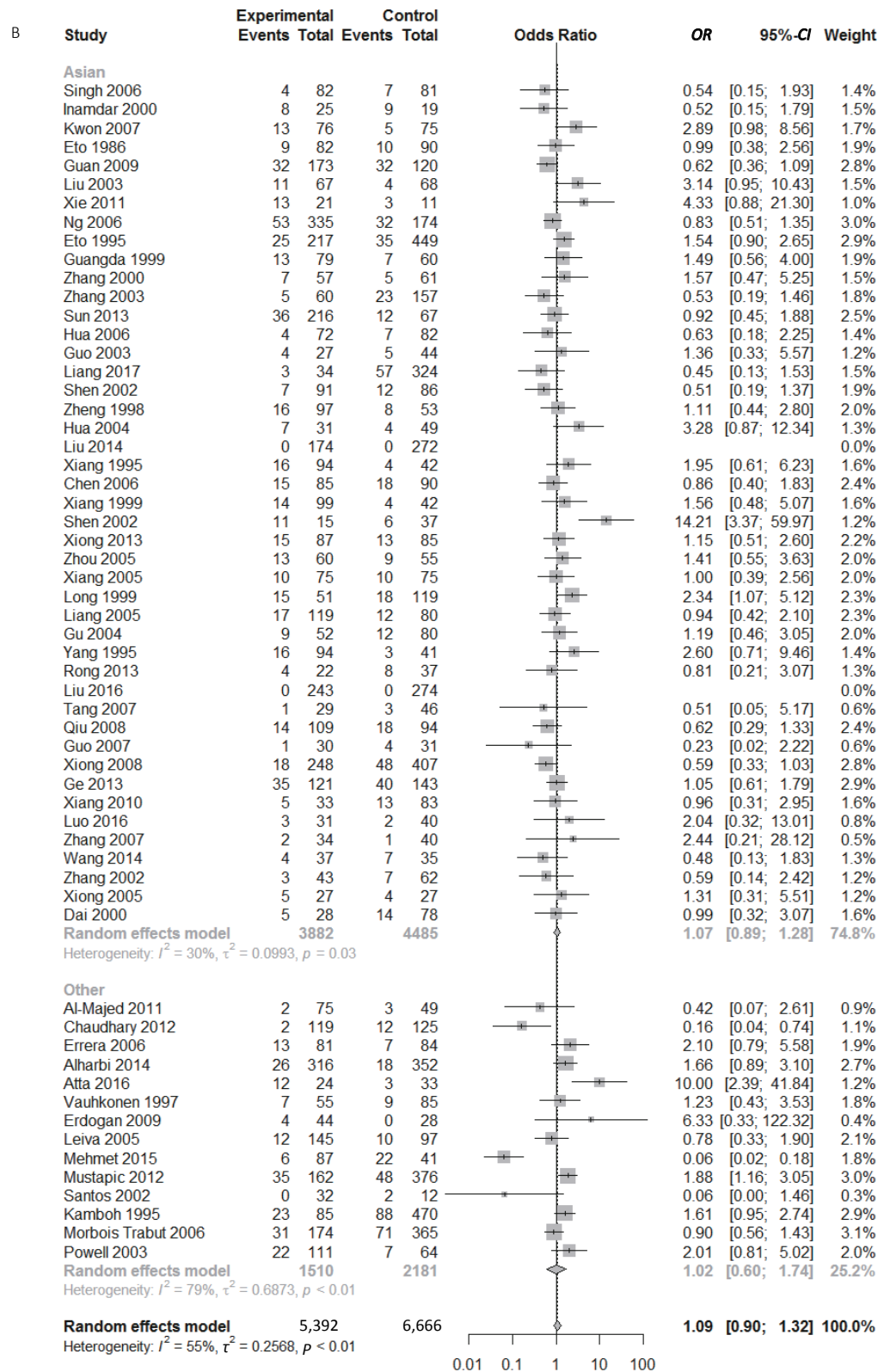
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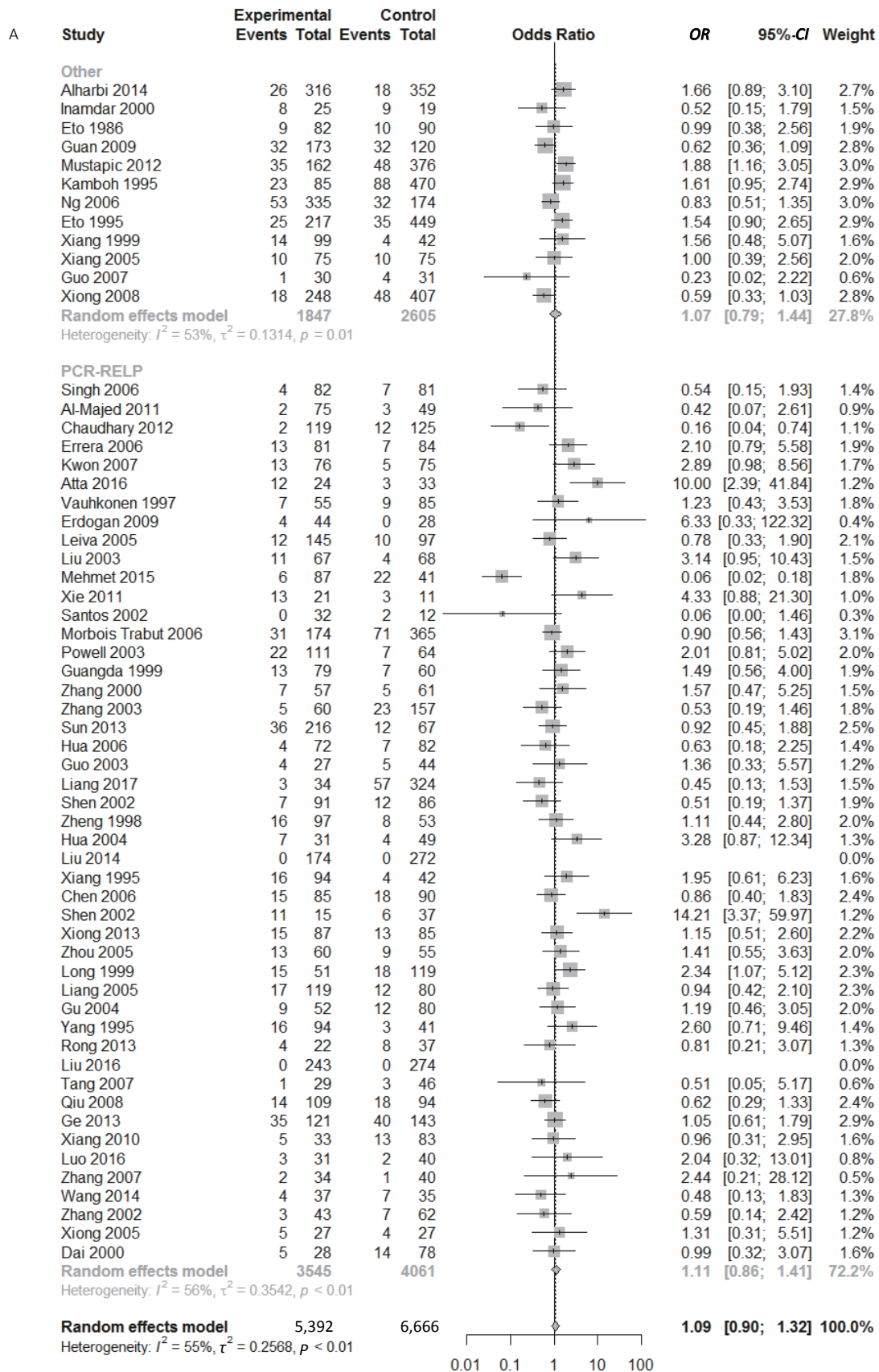


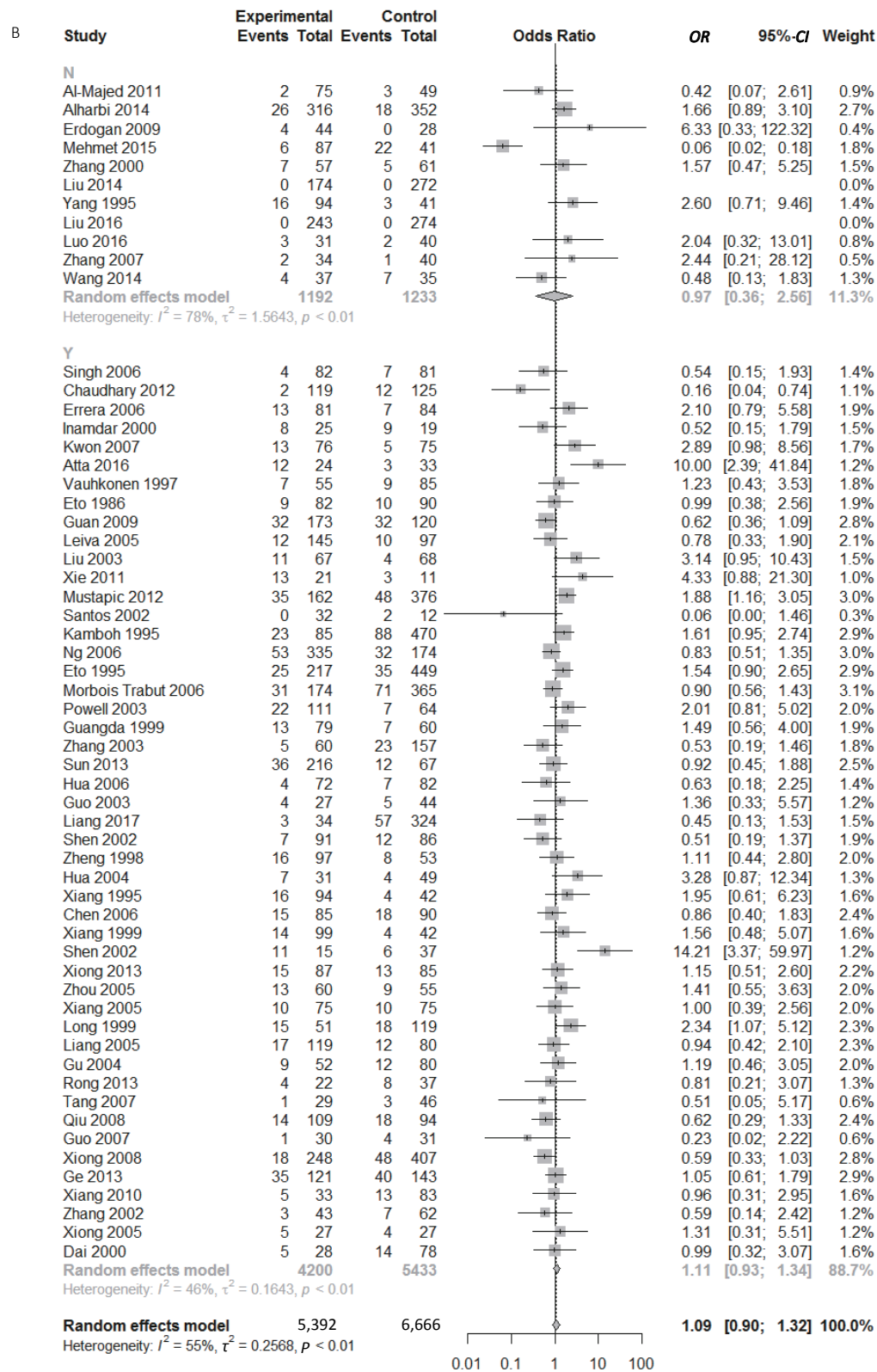
Supplementary Figure S1. (A) Forest plot for associations between type 2 diabetes and *ApoE* $\epsilon 2$ allele vs. $\epsilon 3$ allele in the subgroup based on ethnicity. (B) Forest plot for associations between type 2 diabetes and *ApoE* $\epsilon 2$ allele vs. $\epsilon 3$ allele in the subgroup based on genotype.



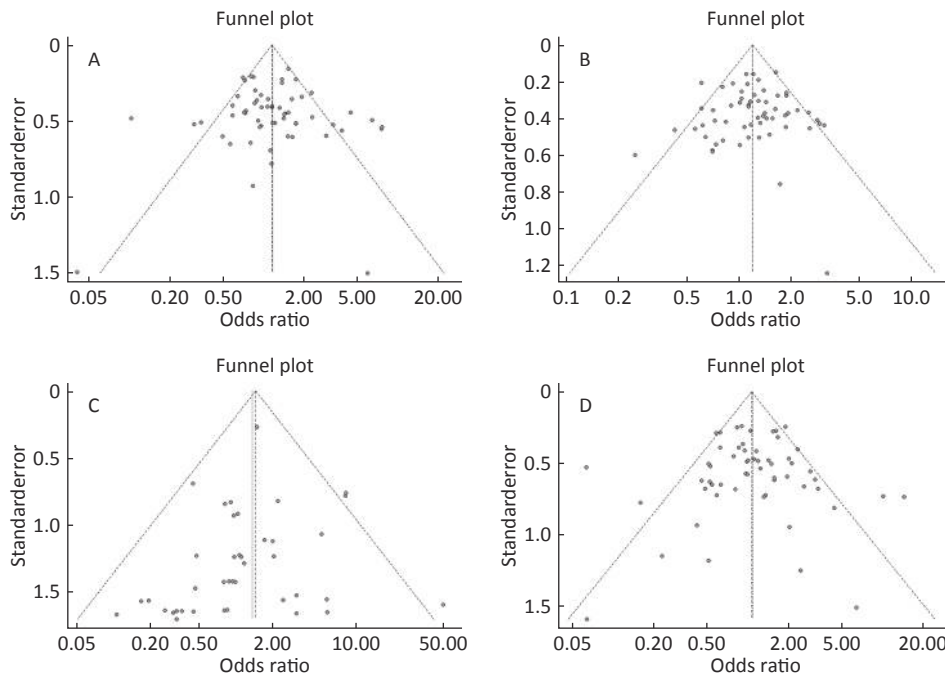


Supplementary Figure S2. (A) Forest plot for associations between type 2 diabetes and *ApoE* $\epsilon 2$ allele vs. $\epsilon 3$ allele in the subgroup based on HWE. (B) Forest plot for associations between type 2 diabetes and *ApoE* $\epsilon 2/\epsilon 3$ genotype vs. $\epsilon 3/\epsilon 3$ genotype in the subgroup based on ethnicity.

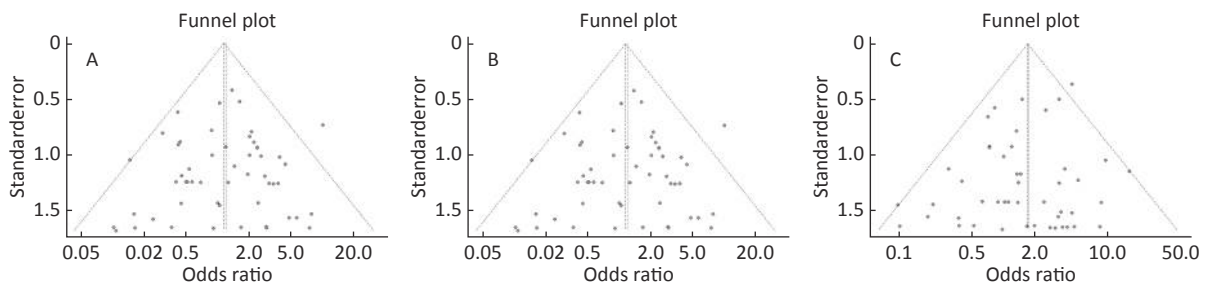




Supplementary Figure S3. (A) Forest plot for associations between type 2 diabetes and *ApoE* $\epsilon 2/\epsilon 3$ genotype vs. $\epsilon 3/\epsilon 3$ genotype in the subgroup based on genotype. (B) Forest plot for associations between type 2 diabetes and *ApoE* $\epsilon 2/\epsilon 3$ genotype vs. $\epsilon 3/\epsilon 3$ genotype in the subgroup based on HWE.



Supplementary Figure S4. (A) Funnel plot of association between type 2 diabetes and *ApoE* $\epsilon 2$ allele vs. $\epsilon 3$ allele. (B) Funnel plot of association between type 2 diabetes and *ApoE* $\epsilon 4$ allele vs. $\epsilon 3$ allele. (C) Funnel plot of association between type 2 diabetes and *ApoE* $\epsilon 2/\epsilon 2$ genotype vs. and $\epsilon 3/\epsilon 3$ genotype. (D) Funnel plot of association between type 2 diabetes and *ApoE* $\epsilon 2/\epsilon 3$ genotype vs. and $\epsilon 3/\epsilon 3$ genotype.



Supplementary Figure S5. (A) Funnel plot of association between type 2 diabetes and *ApoE* $\epsilon 2/\epsilon 4$ genotype vs. and $\epsilon 3/\epsilon 3$ genotype. (B) Funnel plot of association between type 2 diabetes and *ApoE* $\epsilon 3/\epsilon 4$ genotype vs. and $\epsilon 3/\epsilon 3$ genotype. (C) Funnel plot of association between type 2 diabetes and *ApoE* $\epsilon 4/\epsilon 4$ genotype vs. and $\epsilon 3/\epsilon 3$ genotype.