## Letter to the Editor



## The First Reported Case of Co-Infection with *Neisseria Meningitidis* and *Bordetella Pertussis* in the Cerebrospinal Fluid Specimen of a Normal Four-Year-Old Child\*

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Pertussis is an acute and highly contagious respiratory disease caused by Bordetella pertussis (B. pertussis), which occurs in people of all ages and is most dangerous for young children, especially infants. The introduction of whole-cell pertussis vaccines (wPVs) in the 1950s dramatically reduced the incidence of pertussis worldwide<sup>[1]</sup>. However, over the past two decades, many studies have reported the resurgence of pertussis in different countries<sup>[2]</sup>. Epidemiological surveillance in Hubei Province over the last 3 years revealed a clear increasing trend in the incidence of pertussis during the coronavirus disease 2019 (COVID-19) epidemic. B. pertussis mainly invades infant respiratory tracts, triggering acute respiratory infectious diseases. B. pertussis infection can also cause various complications, the most serious being pertussis encephalopathy. However, no obvious change was the observed in **CSF** during encephalopathy, because of which, B. pertussis has rarely been directly detected in the CSF of patients.

Neisseria meningitidis (N. meningitidis) is the chief pathogen responsible for infecting the central nervous system. Most cases of invasive meningococcal disease (IMD) are caused by one of the six meningococcal serogroups: A, B, C, W, X, or Y. Outbreaks of IMD are occurred worldwide, with major serogroups varying geographically and temporally in response to antigenic changes or the use of vaccines. According to surveys between 2010 and 2020, the most prevalent N. meningitidis serogroups in China was NmC (49.7%), followed by NmB (30.2%) and NmW (23.8%), while the most prevalent *N. meningitidis* serogroups in Hubei Province were NmB and NmC and no NmW was reported at present<sup>[3]</sup>.

In this study, we firstly report the co-infection case in China wherein *B. pertussis* and NmW were together detected directly in the CSF of a child. This rare case presented a challenge to our current vaccination strategy in the context of high coverage of *N. meningitidis* and *B. pertussis* containing vaccines.

The patient was a female, 4-years-and-9-months-old, without any recognized immunodeficiency. Before the infection, she had received four doses of acellular pertussis-combined vaccine (DTaP) at 25<sup>th</sup> December 2017, 24<sup>th</sup> January 2018, 26<sup>th</sup> February 2018 and 8<sup>th</sup> April 2019, two doses of group A meningococcal polysaccharide vaccine (MPV-A) at 28<sup>th</sup> March 2018 and 4<sup>th</sup> July 2018, and one dose of group AC meningococcal polys accharide vaccine (MPV-AC) at 2<sup>th</sup> September 2020.

On June 5, the patient had headache and recurrent fever, which lasted for 5 days. The highest axillary temperature was 39 °C and it could be reduced to normal with antipyretics. On the day of onset, the patient had red rash on both lower extremities and two episodes of vomiting. The vomit was a yellow fluid and the abdomen was uncomfortable after vomiting. On Day 5 (June 9), the patient developed frequent cough, expectoration, chills, poor appetite, abdominal pain, and one episode of vomiting. The vomit was non-ejecting,

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constituted by dark-brown stomach contents.

The patient was hospitalized on Day 5. Routine blood tests revealed that the white blood cell count had elevated to  $11.37 \times 10^9/L$  and the percentage of lymphocytes had increased to 68.5%. CSF assays indicated that both karyocytes (930  $\times$  10<sup>6</sup>/L) and red blood cells (300  $\times$  10<sup>6</sup>/L) were detected, the protein content increased to 610 mg/L, and the chloride

**Table 1.** Results of blood testing, CSF assay, q-PCR and nanopore 16S rRNA sequencing

Items	Value in this case	Normal reference value (Children aged 4–5)
Blood testing		
Red blood cells (×10 <sup>12</sup> /L)	4.88	3.9-5.3
White blood cells (×10 <sup>9</sup> /L)	11.37	4.0-10.5
Hemoglobin (g/L)	131	110-150
Lymphocyte (%)	68.5	30–50
Neutrophil (%)	26.0	22–55
Platelet (×10 <sup>9</sup> /L)	305	100-320
CSF assay		
Protein qualitative	Positive	Negative
Karyocyte count (×10 <sup>6</sup> /L)	930	0-8
Red blood cells (×10 <sup>6</sup> /L)	300	0–15
Glucose, mmol/L	2.89	2.8-4.5
Protein, mg/L	610	80-430
Chloride, mmol/L	133.8	120-130
qPCR test based on CSF sam	ple	
N. meningitidis	CtrA Ct = 26.87, sodc Ct = 27.25	Negative
B. pertussis	<i>IS481</i> Ct = 23.11	Negative
N. meningitidis W serogroup	Ct = 34.57	Negative
Nanopore 16S rRNA sequence	cing based on CSF s	ample
N. meningitidis (reads/%)	11,912/72.34	Negative
B. pertussis (reads/%)	2,408/14.62	Negative
Other (reads/%)	2,148/13.04	

**Note**. CSF, cerebrospinal fluid; Mp, mycoplasma pneumoniae; Cp, chlamydia pneumoniae; IgM, immunoglobulin M; IgG, immunoglobulin G. SARS-CoV-2, *C. pneumonia*, *M. pneumonia*, *S. pneumonia*, *S. pyogenes*, *S. agalactiae*, *M. tuberculosis*, and *H. influenza* were not detected by qPCR test. Additionally, Mp-IgM &Mp-IgG, Cp-IgM&Cp-IgG and Influenza A/B Antigen tests were negative. No bacterium was isolated from CSF, blood and sputum samples.

content increased to 133.8 mmol/L (Table 1). Computed tomography (Day 5) and magnetic resonance imaging scans (Day 9 and Day 21) showed that the bilateral cerebral hemispheres were symmetrical, without any obvious abnormal density shadow in the brain parenchyma. The brain cistern, sulcus, and ventricular systems were not dilated (Figure 1B-D). The electroencephalogram showed a slightly slowed background activity. hospitalization, tests for influenza A/B antigens, Mycoplasma pneumoniae and Chlamydia pneumoniae antibodies, and COVID-19 nucleic acids all were negative. No pathogens were detected in CSF, sputum, and blood cultures on Days 5 and 15 (Figure 1A).

On the day of onset, the patient orally took child Ganmao Ning granules, Lianhua Qingwen granules, amoxicillin and clavulanate potassium tablet, and Ambroxol Hydrochloride for 1 day at home. On Days 2-4, the patient was treated with oral and infused oseltamivir and amoxicillin-clavulanate potassium at a local clinic (Figure 1A). On Day 5, the patient was hospitalized and treated with cefotaxime, mannitol, acetylcysteine, and compound ipratropium bromide with budesonide for bronchial and intracranial infections. After 19 days of hospitalization, the patient recovered and was discharged with a of purulent meningitis diagnosis and bronchopneumonitis. The patient reverted to normal health with a return visit on September 27 (Figure 1A).

Later, quantitative PCR performed on CSF samples collected on Day 6 revealed that the patient had been co-infected with N. meningitidis (ctrA Ct = 26.87/sodc Ct = 27.25) and B. pertussis (IS481 Ct = 23.11) (Innowavedx Suzhou Co., Ltd., China, CLRO22A248). The N. meningitidis serogroup was W (Ct = 34.57), determined according to the World Health Organization's protocol (Table 1) (https:// stacks.cdc.gov/view/cdc/11632 and https://www. who.int/publications/i/item/WHO-IVB-14.03). verify the authenticity of the co-infection, full-length bacterial 16S rRNA gene was amplified using BK-16S096 (16sF: CCTTATCATTTAGAGGAAGGAG, 16sR: TCCTCCGCTTATTGATATGC, Hangzhou Technology Co., Ltd., China), followed by nanopore sequencing using the GridION platform (Oxford Nanopore Technologies, UK). Genomic sequencing was then performed on the GridloN sequencing platform (Oxford Nanopore Technologies, UK). The sequencing results were imported into the BAIYI MicroGeno Platform (v 4.1, Hangzhou Baiyi Technology Co., Ltd., [http://www.baiyi-tech.cn/]).

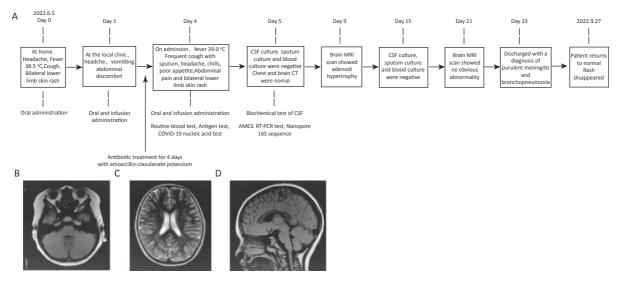
Barcode and adapter sequences were trimmed from the reads using porechop [v0.2.4 https://github. com/rrwick/Porechop]. Seqkit [https://github.com/ shenwei356/seqkit] was used to retain reads between 1.3-1.6k and with an average phred quality score above 12. Subsequently, the filtered sequences were aligned to the MirrOr database using minimap2 [2.6.1 2.24-r1122 https://github. com/lh3/minimap2], retaining only the alignment result for each sequence. The identified species from the matched sequences were recorded as the species of the sequenced reads. A total of 16,468 reads were obtained, of which 72.34% belonged to N. meningitidis and 14.62% to B. pertussis, consistent with the quantitative PCR results. The nanopore sequencing reads have been submitted to the Genome Sequence Archive under accession number CRA008593.

Combined hypoxia is known to encephalopathy in about 0.3% of children with pertussis. Pertussis toxin (PT) can also directly affect pertussis encephalopathy, the fatality rate of which is extremely high, possibly making it a lethal factor for pertussis [4]. B. pertussis is a Gram-negative pathogen usually obtained from respiratory specimens. The few cases of invasive infections caused by В. pertussis mostly involve immunocompromised patients. However, B. pertussis has been detected in the CSF in only a few cases; especially, convincing numbers of B. pertussis reads have been difficult to obtain using highthroughput sequencing methods<sup>[5]</sup>. In this study, more than 2,400 B. pertussis reads were detected in the patient's CSF using full-length 16S rRNA gene nanopore sequencing, providing high-confidence results. Moreover, this process took only 6 hours, from nucleic acid sampling to report.

In previous studies, full-length 16S rRNA gene nanopore sequencing has proven to be a reliable and efficient option for identifying bacterial infections<sup>[6]</sup>. However, this technology lacks well-established guidelines in clinical applications and uniform standards for interpreting the sequencing results. The limitations need to be overcome by researchers in future. Additionally, it could identify multiple pathogens at once and provide more information than traditional RT-PCR test.

The qPCR and nanopore 16S rRNA sequencing test based on CSF sample suggested that this case might involve an N. meningitidis and B. pertussis coinfection. That was also supported by the clinical symptoms. N. meningitidis and B. pertussis infection usually resulted in the symptoms of fever, headache, vomiting, cough and increase of white blood cell count and lymphocytes, which had been found in this case. Moreover, the immune function declines due to the N. meningitidis infection and the action of PT might give *B. pertussis* the opportunity to breach the blood-brain barrier. Similarly, Kugler et al. found that barrier integrity, organelle organization and transmigration of monocytes were affected by pertussis toxin in a human brain microvascular endothelial cell barrier model<sup>[4]</sup>.

Vaccination is considered as the optimal strategy to prevent IMD<sup>[3]</sup>. In this case, the patient had received one dose of MPV-AC. The bivalent MPV-AC



**Figure 1.** The patient's disease progression and MRI examination results. (A) Disease progression flow diagram of the patient; (B–D) Brain MRI scan on Day 9.

vaccine is widely available in China. In addition, the tetravalent MPV-ACWY vaccine has successfully administered for many years, but it is a part of the non-immunization program vaccine management in China, which requires self-payment. Although the development, production, and quality management of MPV-ACWY has greatly progressed in China, it still faces challenges, such as the underestimated burden of IMD the and underdeveloped immunization strategies for meningococcal vaccines.

Over the past decade, the incidence of pertussis has increased in many countries, mostly due to the replacement of whole-cell pertussis vaccine with acellular pertussis vaccine (APV). APV fails to provide long-lasting immunity and establish adequate protection against B. pertussis<sup>[7]</sup>. APV could induce anti-PT immunoglobulin G, and the level induced by the component-purified DTaP-IPV/Hib (Pentaxim, Sanofi) vaccine was significantly higher than that by the co-purified DTaP induced vaccine. Nevertheless, anti-PT immunoglobulin G levels induced by either vaccine decrease over time<sup>[8,9]</sup>. Similarly, the incidence of B. pertussis showed an upward trend in Hubei in recent years according to our previous study<sup>[10]</sup> and unpublished data (epidemiology of pertussis in Hubei Province from 2016 to 2021). It suggested that the current immunization strategies should be further optimized in Hubei province. The need for a booster dose of pertussis-containing vaccine for children entering school and the feasibility of incorporating the tetravalent MPV-ACYW into the immunization program must be investigated. In addition, enhanced active surveillance of IMD and pertussis-associated will enable more targeted vaccine development. Interventions that more appropriate and developed early will help prevent and control NmW infections and pertussis.

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## REFERENCES

- Elomaa A, Advani A, Donnelly D, et al. Population dynamics of Bordetella pertussis in Finland and Sweden, neighbouring countries with different vaccination histories. Vaccine, 2007; 25, 918–26.
- Barkoff AM, Mertsola J, Pierard D, et al. Pertactin-deficient Bordetella pertussis isolates: evidence of increased circulation in Europe, 1998 to 2015. Euro Surveill, 2019; 24, 1700832.
- Xu J, Chen YQ, Yue MM, et al. Prevalence of Neisseria meningitidis serogroups in invasive meningococcal disease in China, 2010-2020: a systematic review and meta-analysis. Hum Vaccin Immunother, 2022; 18, 2071077.
- Kügler S, Böcker K, Heusipp G, et al. Pertussis toxin transiently affects barrier integrity, organelle organization and transmigration of monocytes in a human brain microvascular endothelial cell barrier model. Cell Microbiol, 2007; 9, 619–32.
- Zhang HY, Wang X, Xia H, et al. Metagenomics next-generation sequencing assists in the diagnosis of infant pertussis encephalopathy: a case report. Medicine, 2023; 102, e33080.
- Chen YH, Mao LF, Lai DM, et al. Improved targeting of the 16S rDNA nanopore sequencing method enables rapid pathogen identification in bacterial pneumonia in children. Front Cell Infect Microbiol, 2023; 12, 1001607.
- Le T, Cherry JD, Chang SJ, et al. Immune responses and antibody decay after immunization of adolescents and adults with an acellular pertussis vaccine: the APERT Study. J Infect Dis, 2004; 190, 535–44.
- Zhang ZJZ, Pan JB, Chen M, et al. Seroepidemiology of pertussis in China: a population-based, cross-sectional study. Vaccine, 2021; 39, 1687–92.
- Vidor E, Plotkin SA. Immunogenicity of a two-component (PT&FHA) acellular pertussis vaccine in various combinations. Hum Vaccin, 2008; 4, 328–40.
- He F, Xia X, Nie DW, et al. Respiratory bacterial pathogen spectrum among COVID-19 infected and non–COVID-19 virus infected pneumonia patients. Diagn Microbiol Infect Dis, 2020; 98, 115199.