Effects of Terephthalic Acid on Rat Lipid Metabolism¹

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Objective To study the effect of terephthalic acid (TPA) on lipid metabolism in Sprague-Dawley (SD) rats. **Methods** Five groups of SD rats that ingested 0%, 0.04%, 0.2%, 1%, and 5% TPA, respectively, were included in a 90-day subchronic feeding study. Effects of TPA on levels of serum protein, total cholesterol (TC), triglycerides (TG), high-density lipoprotein cholesterol (HDL), total antioxidative capability (T-AOC), superoxide dismutase (SOD) and malondialdehyde (MDA) were observed. Urine samples were collected and analyzed for concentration of ion. **Results** TPA decreased the level of serum T-AOC in a dose dependent manner. The contents of serum and bladder MDA significantly decreased in 1% and 5% TPA ingestion groups. Serum CuZn superoxide dismutase (CuZnSOD) lowered in groups of 0.2%, 1%, and 5% TPA. TPA subchronic feeding had no significant influences on serum TC, LDL or HDL, but increased serum TG, TP and ALB after administration of 0.04% and/or 0.2% TPA. Concentrations of urinary Ca^{2+} , Mg^{2+} , Na^+ , and K^+ were elevated in 1% and 5% TPA groups. **Conclusion** Antioxidative potential decreased after TPA exposure. MDA increase in serum and bladder tissues was one of the most important reactions in rats which could protect themselves against TPA impairment. The decrease of serum CuZnSOD was related to the excretion of Zn^{2+} .

Key words: Terephthalic acid; SD rat; Serum; Bladder; SOD; MDA; T-AOC; Lipoprotein

INTRODUCTION

Since McCord et al.^[1] reported that superoxide dismutase (SOD) is a marker of oxidative injury, it has been found that mechanism of oxidative stress is not only related with many diseases such as cancer, Alzheimer's disease, familial amyotrophic lateral sclerosis (FALS), and several neurological disorders^[2-4], but also involves toxic reactions of agricultural chemicals, petrochemicals and other compounds^[5-8]. Terephthalic acid (TPA), which belongs to non-genotoxic compounds, is extensively used in the world. Animal studies^[9-10] showed that 3%-5% TPA exposure induced bladder stone within nearly two weeks, followed by bladder hyperplasia and ultimately carcinomas after 90-day subchronic and 2-year chronic feeding studies respectively. Epidemiologic investigations^[11-15] demonstrated that changes in serum angiotensin-converting enzyme, pulmonary function and ventilation, and urinary ions among occupational workers are significantly related

with TPA exposure. To investigate the effect of TPA on the oxidative defense system and elucidate its mechanism, we detected urinary ions, serum lipoprotein and T-AOC, and the content of SOD and malondialdehyde (MDA) in serum and bladder samples from rats which had been exposed to TPA for 90 days.

MATERIALS AND METHODS

Chemicals

Terephthalic acid was obtained from Yi Zheng Chemical Fiber Co. (Jiangsu, China). Total antioxidative capability (T-AOC), superoxide dismutase (SOD) and malondialdehyde (MDA) diagnostic agents were purchased from Nanjing Jiancheng Bioengineering Institute (Jiangsu, China). Serum total protein (TP), albumin (ALB), globulin (GP), total cholesterol (TC), triglycerin (TG), and high-density lipoprotein (HDL) diagnostic agents

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were obtained from Nanjing Kehua Biochemical Co. Ltd. All other chemicals used were commercial products of the highest purity.

Animals and Treatments

Male and female Sprague-Dawley (SD) rats were purchased from Shanghai Animal Center (Shanghai, China), and quarantined for 7 days before the experiments. Room temperature and relative humidity were controlled at $22^{\circ}C\pm 3^{\circ}C$ and $60\%\pm$ 10%, respectively. Fluorescent lighting was provided in a 12 h light/dark cycle. The animals were randomly divided into five groups including 0%, 0.04%, 0.2%, 1%, and 5% TPA, and weighed every week. The treatment was continued for 90 days.

Sample Collection and Storage

All rats were sacrificed on the 90th day. Blood was collected from the retro-orbital venous plexus. After having clotted for 2 h at room temperature, serum were removed and stored at -20 °C. Bladder tissues were weighed, and stored at -70 °C. Fresh urine (100-1000 μ L) was collected between 7:00 and 9:00 a.m. on the day before the animals were sacrificed and stored at -20 °C.

Urinary Ions

Urine Ca^{2+} , Mg^{2+} , Zn^{2+} , Na^+ , and K^+ were analyzed with an AA-6501F atomic absorption flame emission spectrophotometer (Japan) and calibrated with creatinine that was analyzed by a Lisa-500 automatic analyzer (French).

Serum Protein and Lipoprotein

Serum protein, total cholesterol (TC), triglycerin (TG), and high-density lipoprotein (HDL) were measured with a Lisa-500 automatic analyzer (French).

T-AOC, SOD, and MDA Measurement

Bladder was cut into small pieces, homogenized

(1 g/mL) in pre-chilled Tris-HCl buffer (Tris-HCl 0.01 mol/L, EDTA-2Na 0.0001 mol/L, Glucose 0.01 mol/L, and NaCl 0.1 mol/L, pH 7.4) in a tissue homogenizer, then centrifuged at $3000 \times g$ for 15 min at 4°C, and the collected supernatant was assayed immediately. The content of tissue protein was determined by Coomassie brilliant blue G₂₅₀. Levels of T-AOC, SOD, and MDA in bladder and serum were measured according to the product instructions, and their absorbencies were measured by a Beckman DU650 spectrophotometer (USA).

Statistical Analysis

Student's *t*-test was employed to calculate the significance of difference between control and experimental values. P < 0.05 was considered statistically significant.

RESULTS

Body Weight Gain and Urinary Ions

The mean body weight gains of SD rats in all experimental groups had no significant change (Table 1). Levels of urinary Ca^{2+} , Mg^{2+} , Zn^{2+} , Na^+ , and K^+ increased in TPA treatment rats in a dose dependent manner (Table 2).

Serum Protein and Lipoprotein

Serum TP, ALB, and TG increased in TPA treatment groups respectively. Levels of HDL, LDL, and TC had no marked changes in all experimental groups (Table 3).

Levels of T-AOC, SOD, and MDA

TPA decreased serum T-AOC and CuZnSOD in a dose dependent manner (Fig. 1). The mean content of MDA also decreased in bladder and serum after TPA exposure (Figs. 1 and 2).

Group	Male				Female				
	n	Initial Weight	Final Weight	Weight Gain	n	Initial Weight	Final Weight	Weight Gain	
Control	12	138 ± 25	369 ± 42	231 ± 30	12	118 ± 11	254 ± 17	135 ± 18	
TPA 0.04%	12	131 ± 13	361 ± 49	230 ± 41	12	114 ± 12	249 ± 30	135 ± 28	
0.2%	11	132 ± 17	370 ± 37	238 ± 33	12	113 ± 13	266 ± 25	153 ± 29	
1%	12	128 ± 14	356 ± 37	228 ± 33	12	119 ± 11	246 ± 21	127 ± 24	
5%	12	135 ± 15	343 ± 51	208 ± 45	12	119±9	249 ± 19	130 ± 15	

TABLE 1	
Changes in Body Weight (g) and Weight Gain (g) in SD Rats After 90-day Subchronic Feeding of TPA	$(\overline{x} \pm s)$

TABLE 2

Changes of Urinary ion in SD Rats After 90-day Subchronic Feeding of TPA ($\overline{x} \pm s$)

	n	Ca ²⁺ (g/mol Cr)	Mg ²⁺ (g/mol Cr)	Zn ²⁺ (g/mol Cr)	K ⁺ (g/mmol Cr)	Na ⁺ (g/mmol Cr)
Control	23	4.50±0.75	20.92±2.64	0.03 ± 0.00	0.19±0.02	0.38±0.05
TPA 0.04%	24	5.07 ± 0.80	37.41±4.49	0.12 ± 0.01	0.34 ± 0.02	0.49±0.06
0.2%	24	5.23±0.91	44.46±5.75*	0.11±0.01	0.25 ± 0.03	0.29±0.04
1%	24	17.08±2.27**	46.66±5.51**	$0.18{\pm}0.02^{*}$	$0.49{\pm}0.06^{**}$	0.40±0.16
5%	17	84.34±22.02**	103.25±24.64**	0.48±0.13**	$0.32 \pm 0.08^{**}$	$0.80 \pm 0.20^{*}$

Note. Compared with control, *P<0.05, **P<0.01.

TABLE 3

Changes in Serum Protein of SD Rats After 90-day Subchronic Feeding of TPA ($\overline{x} \pm s$	s)
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	п	Cr (µmol/L)	TP (g/L)	ALB (g/L)	GP (g/L)	A/G	TC (mmol/L)	TG (mmol/L)	HDL (mmol/L)
Control	12	38.93 ± 5.90	64.62 ± 4.88	31.08 ± 2.32	33.54 ± 3.07	0.93 ± 0.07	1.20 ± 0.36	0.42 ± 0.10	0.82 ± 0.26
TPA 0.04%	12	35.93 ± 5.46	$69.12\!\pm\!4.95^*$	$34.47 \pm 2.30^{**}$	34.65 ± 3.84	1.00 ± 0.13	1.34 ± 0.33	0.63 ± 0.26	0.96 ± 0.22
0.2%	12	39.68 ± 4.88	67.63 ± 4.57	34.71±1.34**	32.93 ± 3.67	$1.06\!\pm\!0.11^{**}$	1.20 ± 0.25	$0.88\!\pm\!0.43^{**}$	0.89 ± 0.18
1%	12	34.80 ± 5.75	61.18±5.77	30.13 ± 2.51	31.05 ± 3.67	0.98 ± 0.08	1.23 ± 0.41	0.59 ± 0.32	0.95 ± 0.30
5%	12	42.97 ± 10.82	62.54 ± 8.74	30.46 ± 2.92	32.09 ± 6.26	0.97 ± 0.13	1.25 ± 0.41	0.43 ± 0.21	0.89 ± 0.24

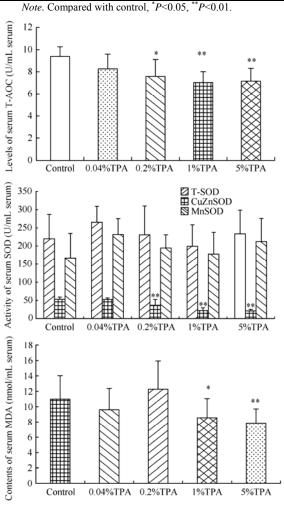


FIG. 1. Effects of TPA on levels of serum T-AOC, SOD, and MDA. (x̄±s) compared with control, *P<0.05, **P<0.01.</p>

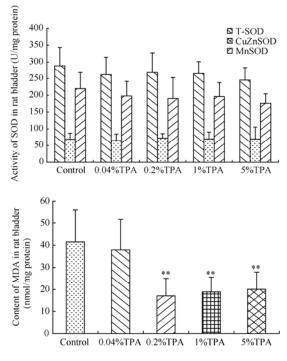


FIG. 2. Effects of TPA on levels of SOD and MDA in rat bladder. $(n=8, \overline{x} \pm s)$ compared with control, ^{**}*P*<0.01.

DISCUSSION

TPA, a kind of organic industrial chemical, is extensively used worldwide. The manifestations of toxicity in baby *ctenopharyngodon idellus*^[16] and the induction of bladder stone, epithelial cell proliferation and ultimately carcinomas by TPA as a non-genotoxic chemical has aroused deep concern over the $world^{[9-10]}$.

Oxidative stress is a self-defense system protecting body against the damage of free radicals, and its dysfunction is always observed in diseases such as tumor and during chemical exposure^[2-7]. The present study illustrated that TPA affected neither the body weight nor weight gain in rats after 90-day subchronic feeding, but it decreased serum T-AOC in a dose dependent manner, representing the total antioxidative capability of the body. In addition, TPA significantly decreased contents of MDA in both serum and bladder tissues, especially after 5% TPA ingestion, suggesting that oxidative defense system initiates to protect the body from TPA induced toxic effects^[17-18]. Our data also showed that the concentration of serum CuZnSOD decreased after TPA treatment. CuZnSOD was distributed in cellular membranes, while the circulating enzymes derived from both hemolysis and peripheral tissues were transported by HDL and/or LDL^[19]. Hall et al.^[20] reported that TPA 20 mg/kg/d decreased serum cholesterol and triglyceride levels in rats. Hence, decreased serum CuZnSOD might be the result of the effects of TPA on lipid metabolism. To elucidate this mechanism, we detected serum protein and lipoprotein in rats after 90-day feeding of TPA. The results have shown that except for TP, ALB, and TG increase in 0.04% and/or 0.2% TPA groups, levels of HDL and LDL have no difference compared with the control. However, TPA treatment significantly increases the concentration of urinary Ca²⁺, Mg²⁺ Zn²⁺, Na⁺, and K⁺. Thus, decreased serum CuZnSOD is related to the increased excretion of Zn^{2+} .

REFERENCES

- McCord J M, Fridovich I (1969). Superoxide dismutase. An enxymatic function for erythrocuprein (hemocurprein). J Biol Chem 244(22), 6049-6055.
- Lin Y, Kikuchi S, Obata Y, et al. (2002). Serum copper/zinc superoxide dismutase (CuZnSOD) and gastric cancer risk: a case-control study. Jpn J Cancer Res 93(10), 1071-1075.
- Yasuda M, Takesue F, Inutsuka S, et al. (2002). Prognostic significance of serum superoxide dismutase activity in patients with gastric cancer. *Gastric Cancer* 5(3), 148-153.
- 4. Noor R, Mittal I J (2002). Superoxide dismutase-applications

and relevance to human disease. Med Sci Monit 8(9), 210-215.

- Panemagalore M, Dowin H A, Byers M E (1999). Occupational exposure to agriculture chemicals: effects on the activities of some enzymes in the blood of farm workers. *Int Arch Occup Environ Health* 72(2), 84-88.
- Georgieva T, Michailova A, Panev T, et al. (2002). Possibilities to control the health risk of petrochemical workers. Arch Occup Environ Health 75(Suppl 1), 21-26.
- Cheung C C, Zheng G J, Lam P K, *et al.* (2002). Relationships between tissue concentrations of chlorinated hydrocarbons (polychlorinated biphenyls and chlorinated pesticdes) and antioxidative responses of marine mussels, perna viridis. *Mar Pollut Bull* 45(1-12), 181-191.
- Kelly S A, Havrilla C M, Brady T C, *et al.* (1998). Oxidative stress in toxicology: established mammalian and emerging piscine model systems. *Environ Health Persp* 106(7), 375-384.
- Heck H D, Tyl R W (1985). The induction of bladder stones by terephthalic acid, dimethyl terephthalate, and melamine (2, 4, 6-triamino-s-triazine) and its relevance to risk assessment. *Regul Toxicol Pharmacol* 5(3), 294-313.
- 10.Chin T Y, Tyl R W, Popp J A, et al. (1981). Chemical urolithiasis. I. Characteristics of bladder stone induction by terepthalic acid and dimethyl terephthalate in weanling Fisher-344 rats. *Toxicol Appl Pharmacol* 58(2), 307-321.
- 11.Wu Z H, Cao J Z, Zhu D M, *et al.* (1998). Studies on relationship between dose of exposure to terephthalic acid dust and pulmonary ventilation function. *Chin J Ind Med* **11**(6), 337-340.
- 12.Li Z, Zhang C, Wang K, *et al.* (1999). Changes in the pulmonary function of factory workers exposure to terephthalic acid. *J Hyg Res* **28**(1), 1-3.
- 13.Gong N, Xu X K, Wang X R, et al. (1999). Changes in urine levels in workers exposed in terephthalic acid. Chin J Ind Med 12(4), 203-208.
- 14.Gong N, He D N, Wang X R, et al. (1999). Study on the changes of urinary biochemical response in workers exposed to terephthalic acid. Chin J Ind Hyg Occup Dis 17(5), 268-271.
- 15.Gong N, He D N, Xu J H, et al. (1999). Effect of terephthalic acid on serum angiotensin-converting enzyme among the occupational workers. *Chin Pub Health* **15**(2), 122-123.
- 16.Gao X (1999). Acute toxicity of PTA of ctenoph-aryngodon idellus. Environ Monit Chin 15(1), 57-58.
- Saran M, Summer K H (1999). Assaying for hydroxyl radicals: hydroxylated terephthalate is a superior fluorescene marker than hydroxylated benzoate. *Free Rad Res* 31(5), 429-436.
- Schimmel M, Bauer G (2002). Proapoptotic and redox state-related signaling of reactive oxygen species generated by transformed fibroblasts. *Oncogene* 21(38), 5886-5896.
- Mondola P, Bifulco M, Seru R, et al. (2000). Presence of CuZn superoxide dismutase in human serum lipoproteins. FEBS Lett 467(1), 57-60.
- 20.Hall I H, Wong O T, Reynolds D J, et al. (1993). Terephthalic acid in Sprague-Dawley rats as a hypolipidemic agent. Arch Pharm 326(1), 5-13.

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