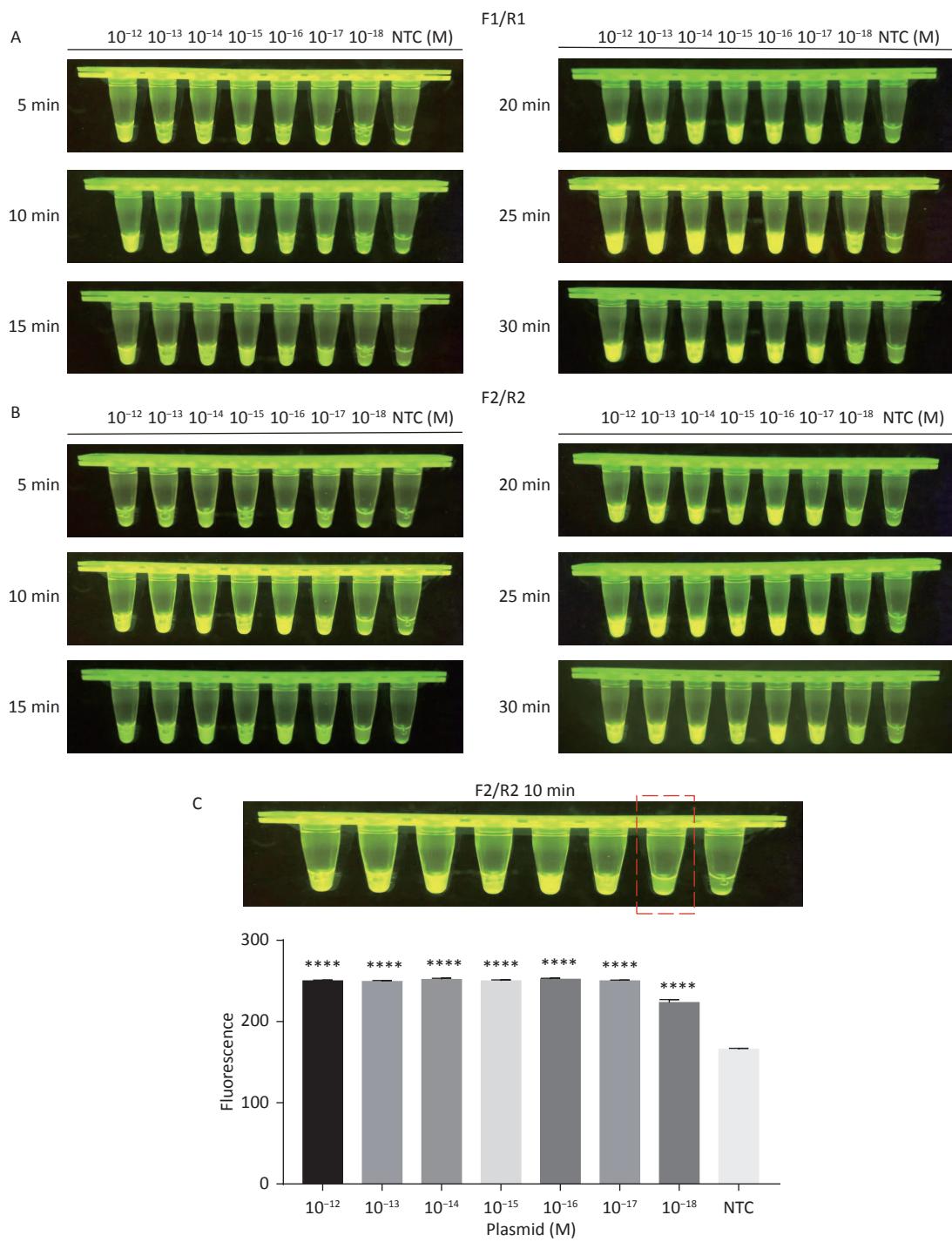
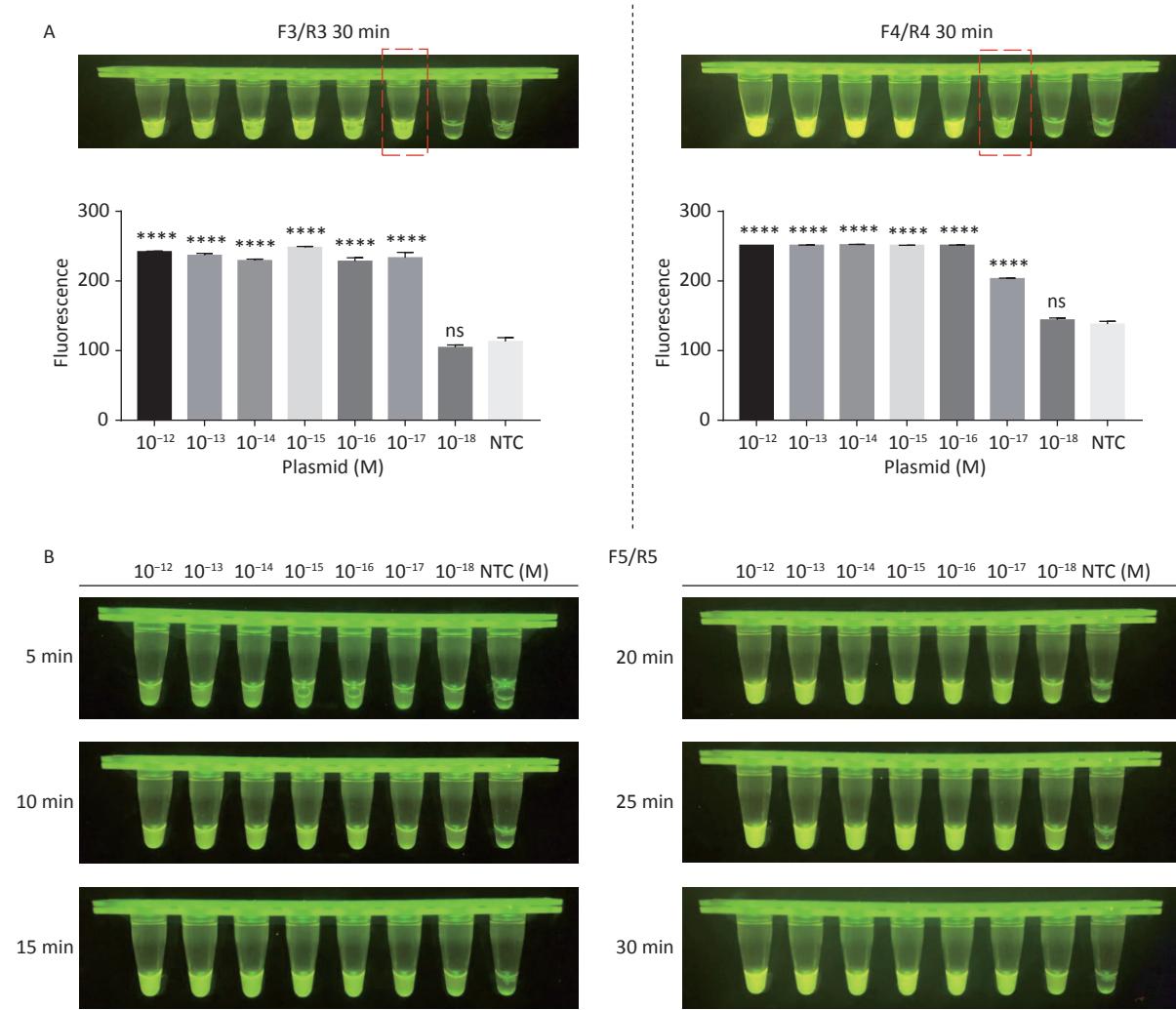




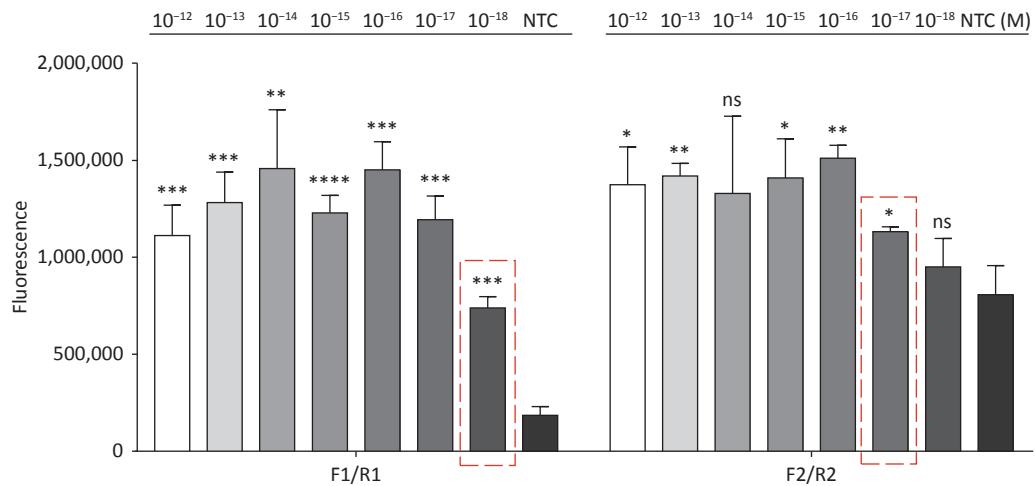
Supplementary Figure S1. Sequence alignment of *tdh* and *trh* genes from 20 *V. parahaemolyticus* genotypes: (A) *tdh* genes (B) *trh* genes. The conserved sequences targeted by crRNA were highlighted with yellow color.



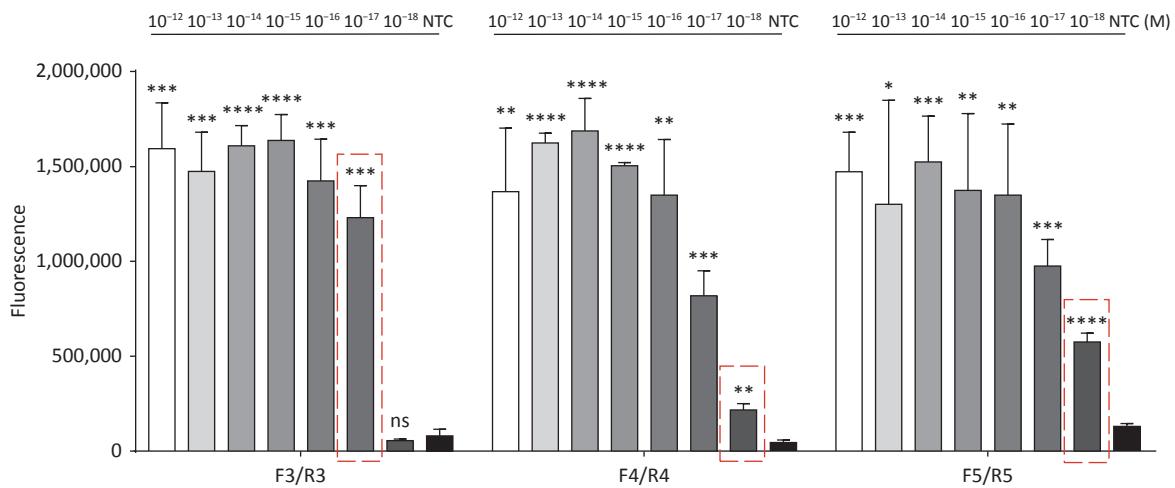
Supplementary Figure S2. Sensitivity of the CRISPR/Cas12a-VD method for detecting the *tdh* gene from 10⁻¹² M to 10⁻¹⁸ M. (A) Amplified products obtained using F1/R1 were added to the Cas12a-mediated cleavage system and incubated for 5–30 min. (B) Amplified products obtained using F2/R2 were added to the Cas12a-mediated cleavage system and incubated for 5–30 min. (C) Amplified products obtained using F2/R2 were added to the Cas12a-mediated cleavage system and incubated for 10 min. The green fluorescent signal was quantified by ImageJ. $n = 3$, two-tailed Student's *t* test; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$ and ns not significant; NTC nontarget control; bars represent mean \pm SD.



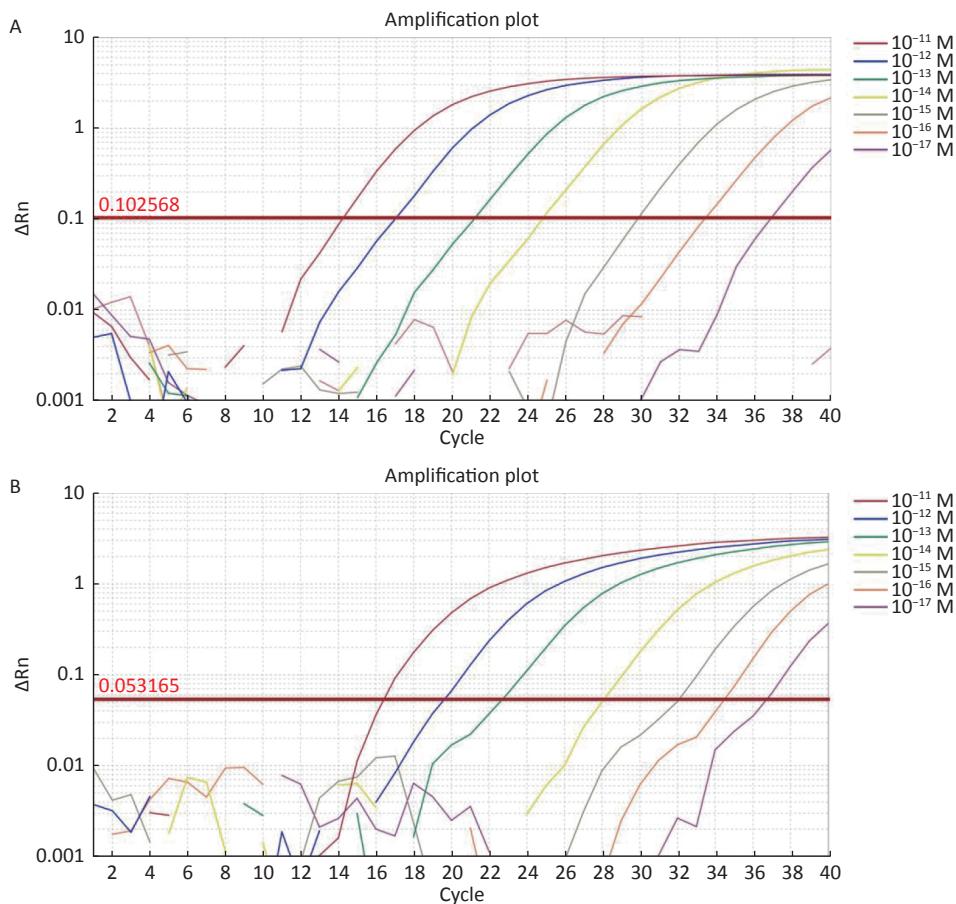
Supplementary Figure S3. Sensitivity of the CRISPR/Cas12a-VD method for detecting the *trh* gene from 10^{-12} M to 10^{-18} M. (A) Amplified products obtained using F3/R3 and F4/R4 were added to the Cas12a-mediated cleavage system and incubated for 30 min. The green fluorescent signal was quantified with ImageJ. (B) Amplified products obtained using F5/R5 were added to the Cas12a-mediated cleavage system and incubated for 5–30 min. $n = 3$, two-tailed Student's *t* test; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$ and ns not significant; NTC nontarget control; bars represent mean \pm SD.



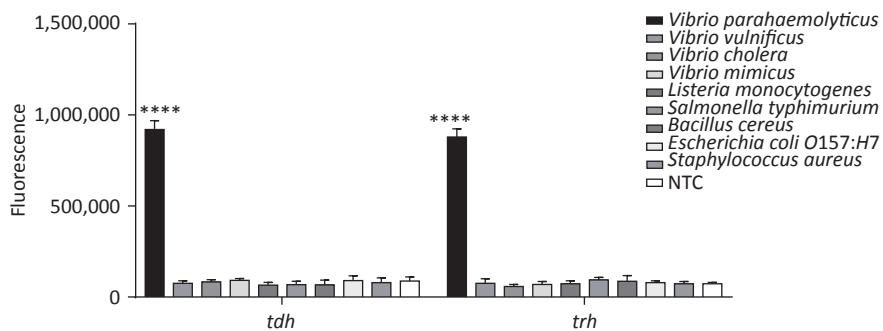
Supplementary Figure S4. Sensitivity of the CRISPR/Cas12a coupled with fluorescence readout for detecting the *tdh* gene from 10⁻¹² M to 10⁻¹⁸ M using primer pairs F1/R1 and F2/R2. *n* = 3, two-tailed Student's *t* test; * *P* < 0.05, ** *P* < 0.01, *** *P* < 0.001, **** *P* < 0.0001 and ns not significant; NTC nontarget control; bars represent mean ± SD.



Supplementary Figure S5. Sensitivity of the CRISPR/Cas12a coupled with fluorescence readout for detecting the *trh* gene from 10⁻¹² M to 10⁻¹⁸ M using primer pairs F3/R3, F4/R4, and F5/R5. *n* = 3, two-tailed Student's *t* test; * *P* < 0.05, ** *P* < 0.01, *** *P* < 0.001, **** *P* < 0.0001 and ns not significant; NTC nontarget control; bars represent mean ± SD.



Supplementary Figure S6. (A) The sensitivity of Real-time PCR to detect *tdh* gene from 10^{-11} M to 10^{-17} M.
(B) The sensitivity of Real-time PCR to detect *trh* gene from 10^{-11} M to 10^{-17} M.



Supplementary Figure S7. Specificity of the CRISPR/Cas12a coupled with fluorescence readout for detecting *tdh* and *trh* genes. $n = 3$, two-tailed Student's *t* test; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$ and ns not significant; NTC, nontarget control; bars represent mean \pm SD.

Supplementary Table S1. Oligonucleotides used to construct plasmid templates

Name	Sequence (5' to 3')
<i>tdh</i> gene fragment	TATTTGCAAAAAAATCATTTTATTTATCCATGTTGGCTG CATTAAAACATCTGTTTGAGCTTCATCTGCCCTTTCC TGCCCCCGGTTCTGATGAGATATTGTTGTTGAGATAC AACTTTAACCCAAGCTCCGGTCAATGTAAAGGTCTGGA CTTTGGACAAACCGTAATGTAAAAAGAAAACCGTAGAAG ATGTTTATGGTCAATCAGTATTACAACGTCAGGTACTAAAT GGTTGACATCTACATGACTGTGAACATTAATGATAAAGACT ATACAATGGCAGCGGTGTTGGCTATAAGAGCGGTATTCTG CTGTGTTGTAAAATCAGATCAAGTACAACATTCAACATTCT ATAATTCTGTAGCTAACATTGGTGAAGATGAAGGTTCTA TTCCAAGTAAAATGATTGGATGAAACTCCAGAATATTG TTAATGTAGAACATGTAGAGGTGTTGAAATATATTG GTAATGTGTATCCAACAAAGAACATGTTTTGAATGT
<i>trh</i> gene fragment	ATGAAACTAAAACCTACTTTGCTTCAGTTGCTATTGGTT CAATGTTTCAGTATCTAAATCATTGCGATGATCTGCCATC AATACCTTTCTCTCCAGGTCGGCTGAACGTATTGTT GTTAGAAAATCAAACAATCAAACGTAATCCCCGGTTAAGGCA ATTGTGGAGGACTATTGGACAAACCGAAACATAAAAAGAAAA CCATACAAAGATGTACGGTCAATCGGTTTACAACAGCAG GTTCAAAGTGGTTAAGCGCTATGACAGTAAACATCAATGG TCATAACTATAGATGGCAGCTTCTGGTTAAAGATGGT ATTCTACGGTCTCACAAATCAGAGAAAACAAGCTAAAGC AAGACTATTCTCGGTAAGCTTTGTTGATGACAGCGAAGA ATCAATACCAAGTATAACTTATTTAGATGAAACATCAGAATAC TTGTTACTGTGCGAGGCATATGAGAGCGGCAATGGACATATGT TCACAAATTAA

Supplementary Table S2. Primers, CrRNA, and SsDNA reporter used in this study

Name	Sequence (5' to 3')
<i>tdh</i> -RPA-F1	CAACTTTAATACCAAGCTCCGGTCAATG
<i>tdh</i> -RPA-R1	CAGCAGAATGACCGCTTATAGCCAG
<i>tdh</i> -RPA-F2	CCGGTTCTGATGAGATATTGTTGTTTC
<i>tdh</i> -RPA-R2	GCAGAAATGACCGCTCTTAGCCAGACACC
<i>trh</i> -RPA-F3	CAATTGTGGAGGACTATTGGACAAACGAAAC
<i>trh</i> -RPA-R3	CTTTAGGCTTGTCTCTGATTTGTGAAGACC
<i>trh</i> -RPA-F4	GTGGAGGACTATTGGACAAACGAAACATA
<i>trh</i> -RPA-R4	TACCGAGGAATAGTCTGCTTAGGCTTGT
<i>trh</i> -RPA-F5	GTGGAGGACTATTGGACAAACGAAACATA
<i>trh</i> -RPA-R5	GACTTTACCGAGGAATAGTCTGCTTAGG
<i>tdh</i> -q-F	AAACATCTGCTTTGAGCTTCCA
<i>tdh</i> -q-R	CTCGAACAAACAATATCTCATCAG
<i>tdh</i> -q-P	FAM-TGTCCCTTCTGCCCGG-TAMRA
<i>trh</i> -q-F	ATCCCGATTGATCTGCCAT
<i>trh</i> -q-R	ATAGCCTCCACAATTGCCT
<i>trh</i> -q-P	FAM-CCTTCTCCAGGTTGGCTGAAGTGT-TAMRA
<i>tdh</i> -crRNA	UAAAAUUUCUACUAAGUGUAGAUUGGUCAAUCAGUAUUCACAA
<i>trh</i> -crRNA	UAAAAUUUCUACUAAGUGUAGAUUGGUUAUAAAAGAUGGUUUU
ssDNA reporter	FAM-TTATT-BHQ1