Review



Research Progress on the Association between Schizophrenia and *Toxoplasma gondii* Infection*

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Abstract: Toxoplasma gondii (T. gondii or Tg), is an obligatory intracellular parasite with humans as its intermediate hosts. In recent years, significant correlations between T. gondii infection and schizophrenia have been reported, including the possible mediating mechanisms. Currently, mechanisms and hypotheses focus on central neurotransmitters, immunity, neuroinflammation, and epigenetics; however, the exact underlying mechanisms remain unclear. In this article, we review the studies related to T. gondii infection and schizophrenia, particularly the latest research progress. Research on dopamine (DA) and other blood-brain neurotransmitters, the inflammatory factors, disease heterogeneity, and other confounders is also discussed. In addition, we also summarized the results of some new epidemiological investigations.

Key words: *Toxoplasma gondii*; Schizophrenia; Neurotransmitters; Neuroinflammation; Immunity; Epigenetics

Brief Introduction of Toxoplasma gondii

Toxoplasma gondii is an obligatory intracellular protozoan widely distributed in nature and has a complex life cycle. Almost all endothermic organisms, including humans, are susceptible to *T. gondii* infection, resulting in toxoplasmosis, which can cause major damage. Notably, almost one-third of the global population is chronically infected with *T. gondii* 1. *T. gondii* was first found in the tissues of the North African rodent gondii by Nicolle and Manceaux in 1908^[2]. In the same year, Splendore also found them in the tissues of rabbits. *T. gondii* has five stages in its life cycle: trophozoites (including bradyzoites and tachyzoites),

gametophytes, cysts, schizonts, and oocysts, each with a unique shape.

Humans, pigs, and cows can act as intermediate hosts for T. gondii; however, cats and other felines are definitive hosts. Multiple stages of T. gondii infection could occur, and intermediate hosts can be infected via various routes, such as the ingestion of oocysts, cysts (bradyzoites), or tachyzoites. After penetrating the intermediate host and invading nucleated cells, the parasite prefers to multiply in the liver, lymph nodes, neurological system, and muscular tissue. T. gondii is a single parasitic species; however, different strains exist. The virulence and sensitivity of T. gondii vary when infecting different hosts. Using six polymerase chain reaction and restriction fragment length polymorphism markers, Howe et al. found that most of the 106 Ta strains from North America and Europe belonged to three clonal lineages, namely classical types I, II, and III^[4]. Serotyping studies have shown that type I strains significantly increase the incidence of psychiatric illnesses^[5,6]. In epidemiological studies of *T. gondii* in China, 12 genotypes have been isolated, among which ToxoDB#9 accounted for the highest proportion (65.5%) and was named Chinese1, followed by the classic type I lineage (18.3%)^{1/3}. Notably, the Chinese1 type includes both virulent and attenuated strains, and its pathogenic mechanism requires further study^[8]. These reports suggest that the pathogenic outcomes of T. gondii infection differ in different geographic regions.

Pathogenic Mechanism of Toxoplasma gondii

Most of the time no significant clinical symptoms have been observed after humans become infected with *T. gondii*. When a parasite intracellularly

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invades an immunocompetent patient, the host may be stimulated to build protective immunity and develop tissue cysts^[9]. The host often remains asymptomatic. Most patients with schizophrenia infected by the parasite are such healthy hosts. Immunosuppressed or immunocompromised patients, such as those with acquired immune deficiency syndrome, may experience extensive parasite proliferation and increased pathogenicity from the reactivation of tissue cysts to the tachyzoite stage, resulting in acute toxoplasmosis, which can cause serious pathological injury or even death^[10]. Moreover, the parasite may be transmitted from the placenta to the fetus if the immunocompetent mother is infected during pregnancy or within three months before conception [11]. Blood transfusions (whole blood or white blood cells) or organ transplants can also transmit the parasite if the donor has T. gondii infection.

A growing number of reports have confirmed that central nervous system (CNS) dysfunctions, including toxoplasmic encephalitis, may occur in immunocompetent hosts infected with *T. gondii*^[12-14]. Parasites can migrate to the host brain within seven days of infection. Experimental studies in mouse models of *T. gondii* infection have shown that approximately 10% of neurons and astrocytes and 30% of microglia may be infected with tachyzoites^[15].

Toxoplasmosis poses a serious risk to humans, particularly developing fetuses. Pregnant women infected with *T. gondii* may experience abortion, stillbirth, and intrauterine growth retardation; infants may develop congenital toxoplasmosis, leading to problems in the CNS, vision, and hearing^[16]. In recent decades, there has been increasing interest in the potential connection between *T. gondii* infection and mental diseases. However, the exact mechanism by which this parasite causes mental illnesses remains unknown.

Correlation between T. gondii Infection and Schizophrenia

Schizophrenia is a heterogeneous disorder characterized by the following three features: positive symptoms, such as hallucinations and delusions; negative symptoms, such as disordered thought processes, social withdrawal, and cognitive and mnemonic impairments. Although the exact etiology of schizophrenia remains unknown, it is a serious mental illness affecting 1% of the population. Recent studies have shown that some cases of schizophrenia may be related to *T. gondii* infections.

To date, many researchers have demonstrated this correlation using serological studies, animal models, and other methods. Currently, the most commonly used technique for investigating the connection between *T. gondii* infection and schizophrenia is serological research.

Chen et al. studied psychopaths in the southeastern region of Zhejiang Province, China, using a case-control methodology. Through enzymelinked immunosorbent assays, they found that the seroprevalence of anti-T. gondii IgG and IgM levels significantly higher in patients with schizophrenia than healthy controls^[17]. Similarly, Veleva et al. found that compared to people with schizophrenia who were serum-negative for T. gondii, patients who were positive for T. gondii had more severe damage in a range of cognitive domains, as well as more severe psychopathological damage^[18]. In addition, Grada et al. conducted a case-control study in western Romania and found that patients diagnosed with schizophrenia had a significantly higher seroprevalence of IgG antibodies against *T. gondii* than healthy controls^[19]. Liu et al. conducted a cross-sectional study on the status of T. gondii infection in patients with schizophrenia in Shandong Province, China. They found that the serum positivity rate for T. gondii infection in patients with schizophrenia was relatively high, and it was estimated that age was an influencing factor for this [20]. In recent years, similar studies have been conducted in Jiangsu Province, China^[21].

However, a study by Ademe et al. revealed that the seroprevalence of toxoplasmosis in individuals with schizophrenia was only marginally higher than that in normal individuals, and the statistical data indicated no substantive difference^[22]. Contopoulos et al. conducted a meta-analysis of 66 studies on the association between T. gondii infection and schizophrenia involving 11,540 patients with schizophrenia and 69,491 normal controls. Their findings demonstrated a robust statistical correlation between T. gondii infection and schizophrenia despite the presence of numerous methodological flaws in the included case-control studies^[23]. Other studies have also suggested a strong association between *T. gondii* infection and schizophrenia^[24-28].

Furthermore, an association between *T. gondii* infection and schizophrenia was demonstrated using animal models. Studies on the behavior of *T. gondii*-infected mice were conducted in 1978 to compare changes in their learning capacities^[29]. In Morris water maze experiments, Daniels et al. found that latent toxoplasmosis alters neurocognitive

symptoms, especially memory deficits, in infected rats^[30]. High specificity for cat urine has also been demonstrated in mice with chronic T. gondii infection^[31]. A study by Boillat et al. (2020) showed that infected mice not only have a specific tendency toward the smell of cats but also exhibit decreased fear and aversion to various predators. They also demonstrated that behaviors manipulated by T. gondii included increased activity, impulsiveness, and reduced fear, which correlated positively with the increased number of cysts in the cerebral cortex of the brain^[32]. Rats and mice do not exhibit the entire phenotypic spectrum of schizophrenia. Schizophrenia manifests as a wide range of symptoms that differ across individuals. Although this complexity cannot be fully summarized in rodent models, specific symptom classes can be simulated using behavioral models^[33].

Furthermore, other research demonstrated that Т. gondii infection may only exacerbate schizophrenia rather than cause it. Holub et al. conducted a survey of 251 patients, of whom 77.3% were infected with T. gondii. The positive symptom assessment scores of the infected patients were significantly higher than those of the uninfected patients, whereas the negative scores were equivalent to those of the general population. Particularly in terms of irritability, cognitive disorders, hostility, and behavioral disorders, there were significant differences between infected patients and patients negative for anti-T. gondii antibodies. T. gondii-infected patients frequently exhibit erratic, impulsive, easily agitated, and irritable behavior^[34]. Similarly, a survey by Arjen et al. that included young and patients with mild schizophrenia, patients with anti-T. gondii antibodies had significantly more severe positive symptoms, whereas negative symptoms remained unchanged^[35].

Progress on Elucidating the Mechanism of Toxoplasma Gondii Infection-related Schizophrenia

According to evolutionary theory, some researchers have proposed evolutionary hypotheses, such as the behavioral manipulation effect of parasites, and they contend that *T. gondii* infection will prevent people from entering society, which will aid in the survival of the parasite^[36]. However, it is difficult to verify specific aspects of this hypothesis. Therefore, we should focus on studying molecular biological mechanisms. Presently, the focus of this study was immunological responses, brain encapsulation, central neurotransmitters, and brain

tissue structures. However, an increasing number of researchers have employed recently developed techniques to elucidate the genetic basis of *T. gondii* infection. Additionally, studies on drug use and metabolism have complemented the association between schizophrenia and *T. gondii* infection. Below is a summary of the review considerations regarding the methods mentioned above (Figure 1). Central Neurotransmitter Changes Current research has shown that the occurrence of schizophrenia is closely related to an imbalance in regulating central neurotransmitters, such as DA, glutamate, 5-hydroxytryptamine (5-HT), and gammaaminobutyric acid (GABA); its core hypothesis is the dopamine hypothesis. Therefore, researchers have considered central neurotransmitters, such as dopamine, while examining the connection between T. gondii infection and schizophrenia. As early as 1985, Stibbs found that chronic infection with T. gondii caused a significant increase in DA in the brain of mice, but acutely infected mice had unchanged dopamine levels, elevated homovanulanic acid levels, and decreased norepinephrine levels; serotonin and 5-hydroxyindoleacetic acid levels were not altered in infected mice. These findings offer fresh insights into the connection between T. gondii infection and neurotransmitter systems and imply that T. gondii infection may affect neurotransmitter

Several studies have linked chronic T. gondii infection to increased dopamine release, emotional problems, and aberrant behavior [38,39]. Moreover, some studies have found that drugs targeting the dopamine system can attenuate behavioral abnormalities that occur after *T. gondii* infection^[40]. However, not all forms of schizophrenia can be explained by the dopamine hypothesis. Gaskell et al. used dual-active amino acid hydroxylase as a study subject and knocked out two aromatic amino acid hydroxylase genes, AAH1 and AAH2, in the T. gondii genome and found that the genes could encode levodopa (L-DOPA), a direct precursor protein for dopamine production. Thus, this enzyme plays an important role in *T. gondii* metabolism, growth, and development, suggesting that T. gondii affects DA production and synaptic transmission^[41]. McFarland et al. found that the mechanism by which T. gondii infection affects the dopamine system might not be related to the AAH2 gene, which is not required for T. gondii to induce behavioral abnormalities^[42]. Furthermore, Afonso et al. suggested that T. gondii infection produces behavioral changes in mice, such as aggression. However, the existence or lack of

levels in mice, thereby affecting their behavior^[37].

AaH2 tyrosine hydroxylase activity encoded by the Toxoplasma genome has little bearing on these behavioral alterations^[43]. Dopamine plays a regulatory role in subsequent *T. gondii* infections, and there is an interaction between them. Some researchers have shown that dopamine stimulates the growth and spread of *T. gondii* trophozoites in cultures of neonatal rat astrocytes and infected human fibroblasts. This effect may be related to the capacity of the parasite to influence host cell immune responses and immune clearance^[44].

David et al. examined the impact of T. gondii

infection on glutamate metabolic homeostasis and neuronal function in the CNS. They discovered that *T. gondii* infection inhibits the expression and function of the glutamate transporter protein glutamate transporter-1 (GLT-1) in the CNS, which interferes with the uptake of glutamate by neurons, causing glutamate accumulation in the CNS and neurons and triggering neuronal death and neurotransmitter dysregulation. These results suggest that *T. gondii* infection may inhibit GLT-1 to promote neuronal cell death, causing pathological processes, such as dysregulation of neurotransmitter

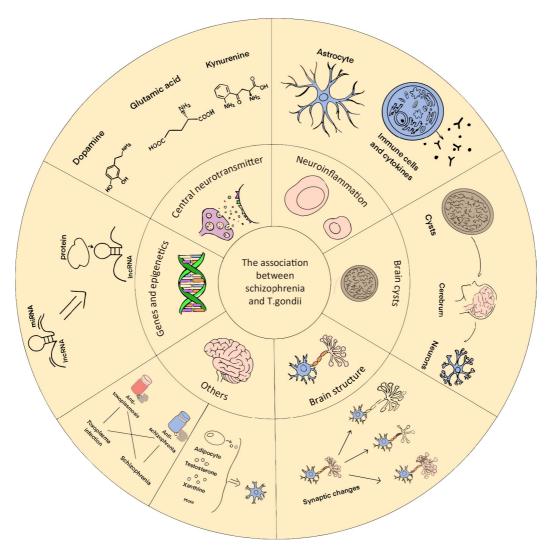


Figure 1. Summary of the association between schizophrenia and *Toxoplasma gondii* infection. This article reviews the association between schizophrenia and *T. gondii* infection from the perspective of molecular biological mechanisms, including (1) central neurotransmitters, (2) neuroinflammatory response and altered immune functions, (3) distribution of *T. gondii* cysts in the brain, (4) structural changes in the neural tissue in the brain, (5) gene regulation and epigenetics, (6) combination therapy (which suggests an association between schizophrenia and *T. gondii* infection), and (7) changes in other metabolic pathways. IncRNA, long noncoding RNA; miRNA, microRNA.

homeostasis and neuronal damage in the CNS^[45]. Acquarone et al. found that mice infected with T. gondii were stunned by the amplitude of the startle, showing reduced glutamate and D-serine levels in the prefrontal cortex and hippocampus. Their results suggest that infection with the T. gondii ME49 strain could induce behavioral changes associated with psychiatric disorders and glutamatergic dysfunction^[46]. neurotransmission Excitatory ionotropic glutamate receptors, including the Nmethyl-D-aspartate receptor (NMDAR), are involved in synapse formation, neuronal development, and other processes. A previous study investigating the mechanism of immune cross-reactions proposed that the NMDA model was connected to the association between *T. gondii* and schizophrenia. Moreover, different NMDAR subunits are targeted at different exposure times, especially NMDA 2D, the target^[47]. potential Glutamate decarboxylated by glutamic acid decarboxylase to produce GABA. Since GABA is the primary inhibitory neurotransmitter in the brain, an imbalance between excitatory and inhibitory neuronal activity could result from a disruption in GABA production. T. gondii infection increases the dispersion of glutamic acid decarboxylase 67. a key GABA synthesis enzyme, in the mouse brain, thereby affecting GABA synthesis^[48].

Tryptophan is reportedly broken down by astrocytes, which subsequently produce a significant quantity of kynurenine (KYNA), which affects peripheral neurons. KYNA is an antagonist of NMDAR and elevated levels of KYNA can block the glutamate recognition sites of NMDAR. Pearce et al. found higher levels of tryptophan in patients with schizophrenia with longer onset and peak latency, contrary to the theory that tryptophan depletion leads to enhanced dopaminergic responses. Furthermore, they showed that the schizophrenia group had lower levels of KYNA in the blood and higher levels of KYNA in their cerebrospinal fluid^[49]. Thus, they hypothesized that KYNA is an important link between T. gondii infection and schizophrenia. In addition, Mahmoud et al. found that T. gondii interferes with the synthesis of 5-HT through the KYNA pathway^[50].

In summary, although there is a large amount of evidence that neurotransmitters such as dopamine and glutamate are strongly associated with schizophrenia and *T. gondii* infection, there are still some issues that need to be addressed. First, more studies on central neurotransmitter regulatory pathways are needed, as some signaling pathways

have not yet developed into a fully functional system. Second, there is a need to investigate why the levels of neurotransmitters, such as dopamine, are elevated after T. gondii infection, for which we can try to start elucidating from the aspects of genetic and epigenetic regulation. Many research methods rely on animal studies; however, it is important investigate how to neurotransmitter alterations in the brain in vivo align with the published results in other models like animal behavioral experiments. An overview of the modifications to the core system is provided below (Table 1).

Neuroinflammatory Response and Altered Immune **Function** T. gondii infection typically triggers a robust immune response, particularly neuroinflammatory response, because T. gondii is viewed as a foreign agent that invades the organism. It is believed that the blood-brain barrier protects the brain and is not easily affected by immune responses. However, T. gondii could infect the brain rapidly and break through the blood-brain barrier. Konradt et al. found that T. gondii replicates in endothelial cells (ECs) and then lyses them, as evidenced by in vivo imaging in acutely infected mice^[51]. Kim et al. constructed an *in vitro* microfluidic model of acute T. gondii infection and transendothelial migration. They found that T. gondii could rapidly infect ECs in the acute phase and then pass through the extracellular matrix to neurons. However, it is difficult to experimentally demonstrate whether tachyzoites directly infect neurons^[52]. Moreover, from the perspective of neuroinflammation, the destruction of the bloodbrain barrier may be an important pathogenic mechanism (Figure 2).

In addition to the acute phase, the chronic phase and latent *T. gondii* infection are also intimately linked to the inflammatory response. Estate et al. found that *T. gondii* infection leads to a reduction in the number, length, and branching of mouse brain capillaries, accompanied by neuroinflammation, microglia activation, and an increase in the number of white blood cells in the brain capillaries. Therefore, it can be inferred that *T. gondii* infection activates neuroinflammation, changes the permeability of the blood-brain barrier, reduces angiogenesis, and causes microcirculation dysfunction. The study of Estate et al. was the first to show that even during the asymptomatic phase of the disease, T. gondii infection causes long-term changes in the cerebral microvascular structure that are linked to endothelial dysfunction and persistent

leukocyte-endothelium interactions, suggesting elevated vascular inflammation^[53]. Hamdani et al. found that patients with schizophrenia who were infected with *T. gondii* were more likely to develop changes in the immune-inflammatory process by testing their antibody and immunoglobulin levels^[54]. Fond et al. discovered that latent *T. gondii* infections were three times more common in patients with schizophrenia than in the controls and that latent

infections in these patients were linked to specific mental symptoms and peripheral inflammation [55]. Moreover, several studies have demonstrated an important association between the schizophrenic course of T. gondii infection and neuroinflammation, from the antibody level to the brain imaging, including cytokines such as interleukin-6 (IL-6), transforming growth factor- β 1 (TGF- β 1), interferongamma (IFN- γ), and various other immune proteins,

Table 1. A brief summary of the results of existing studies on the changes in central neurotransmitters

Year	Author	Target	Main findings [*]
1985	Stibbs et al. ^[37]	Catecholamines and indoleamines	In the acute phase: vanillic acid (\uparrow) , dopamine levels remain unchanged, epinephrine (\downarrow) . In the chronic phase: DA levels (\uparrow) , serotonin levels are not altered.
	Gaskell et al. [41]	L-DOPA and DA	The genome of the protozoan parasite <i>Toxoplasma gondii</i> was found to contain two genes encoding tyrosine hydroxylase that produce L-DOPA. <i>T. gondii</i> AaaH1 is constitutively expressed, while the other gene, <i>T. gondii</i> AaaH2, is induced during the formation of the bradyzoites of the cyst stages of the life cycle. After infection: K ⁺ -induced release of DA (\uparrow). <i>T. gondii</i> orchestrates a significant
2011	Prandovszky et al. ^[40]	DA	increase in DA metabolism in neural cells.
2012	Strobl et al. ^[44]	DA	Higher doses of dopamine significantly increased the number of <i>T. gondii</i> tachyzoites in human fibroblast host cells. DA antagonists had no significant effect on DA-stimulated tachyzoite production in human fibroblasts. DA significantly increased the number of intracellular tachyzoites in primary cultured neonatal rat astrocytes.
2015	Brooks et al. ^[48]	GAD67 and GABA	Seizures and other neurological complications seen in <i>Toxoplasma</i> -infected individuals are due, at least in part, to changes in GABAergic signaling.
2016	Mahmoud et al. ^[50]	Serotonin(5-HT), DA, and NE.	With/without DXM at 60 dpi: DA turnover (个), 5-HT turnover (个). DA and norepinephrine levels are not associated with the observed behavioral changes.
2016	David et al. ^[45]	Glutamate and GLT-1	After infection, the major astrocyte glutamate transporter GLT-1 (\downarrow) , extracellular glutamate concentration (\uparrow) , dendritic spines (\downarrow) , immune reactivity of VGlut1 and NeuN (\downarrow)
2017	Afonso et al. [43]	тн	Parasite-secreted AaaH2 TH is neither necessary for the generation of risky behavior nor for the increased trappability observed during chronic <i>Toxoplasma</i> infection.
2017	Lucchese et al. [47]	NMDAR	The NMDA 2D subunit, which is mainly expressed in parvalbumin-positive interneurons, appears to be a hotspot for potential <i>T. gondii</i> -induced cross-reactive immune attacks.
2018	McFarland et al. ^[42]	DA, DAT and VMAT2	After infection of BALB/c mice: blunted response to amphetamine or cocaine, expression of DAT and VMAT2 (\downarrow). AAH2 is not required for <i>T. gondii</i> infection-produced DA-dependent neurobehavioral abnormalities.
2020	Ibrahim et al. ^[38]	IgG anti- <i>Toxoplasma</i> dopamine	Dopamine levels in +ve IgG schizophrenia are higher than those in -ve IgG schizophrenia. Chronic <i>T. gondii</i> infection can lead to elevated DA levels, which can lead to schizophrenia.
2020	Pearce et al. ^[49]	Kynurenine	KYN pathway markers predict a slowing of startle latency in SCZ subjects and in those with chronic <i>T. gondii</i> infection; this was not seen in CON or <i>T. gondii</i> seronegative subjects.
2022	Acquarone et al. ^[46]	Glutamate and D-serine	Chronic infection: glutamate and D-serine (\downarrow), but behavioral and cognitive aspects are maintained.
2023	Omidian et al. ^[39]	Dopamine	Serum DA on the first to the fourth day after <i>T. gondii</i> inoculation was the same as the negative control, but increased on the fifth day. DA production is low in acute toxoplasmosis; chronic disease increases DA production.

Note. *(↑), upregulation; (↓), downregulation. L-DOPA, levodopa; GABA, gamma-aminobutyric acid; NE, norepinephrine; DXM, dexamethasone; GAD67, glutamate decarboxylase 67; GLT-1, glutamate transporter-1; 5-HT, 5-hydroxytryptamine; DA, dopamine; GLT-1, glutamate transporter-1; TH, Tyrosine hydroxylase; NMDAR, N-methyl-D-aspartate receptor; DAT, dopamine transporter; VMAT2, vesicular monoamine transporter 2; VGlut1, recombinant vesicular glutamate transporter 1; NeuN, neuronal nuclei; KYN, kynurenine; SCZ, schizophrenia; CON, chronic *Toxoplasma gondii* infection.

even complement expressed by astrocytes, microglia, and infiltrating monocytes^[56-59].

Neuroinflammation may also be associated with brain cysts. Using infected C57BL/6 mice as models, Barrios et al. discovered a positive correlation between the number of T. gondii cysts in the brain and the upregulation of TNF and CC-chemokine expression in the brain tissues. Additionally, elevated levels of IFN-y, TNF, and CCL5/MCP-2 in the peripheral blood suggested that the persistence of parasite cysts may result in persistent neuroinflammation^[58]. The presence of parasitic cysts may also lead to the disruption of the bloodbrain barrier, allowing many cytokines to enter the brain tissue. Xiao et al. found that immunotherapies targeting PD-1 pathway alleviated neuroinflammation induced by chronic T. gondii infections and suggested that this was associated with a reduction in the numbers of T. gondii tissue cysts during treatment^[60].

Numerous studies have investigated the various impacts when a mother is infected with *T. gondii* during pregnancy. Romero et al. explored the mechanisms of maternal transmission to the offspring and found that pre-pregnancy exposure to *T. gondii* induces the production of maternal

pathogenic antibodies that recognize and bind to fetal brain protein mimics, leading to neurostructural, cognitive, and social alterations in the offspring ^[61]. According to the observations of early pregnancy infection with *T. gondii* by Bi X. et al., the immune escape of *T. gondii* at the maternal-fetal interface may be linked to the aberrant expression of CD47 and its ligands, signal regulatory protein- alpha (SIRP α) and thrombin-sensitive protein 1 (TSP-1), in the mother and fetal mice during mid-to-late gestation ^[62].

In conclusion, the prevailing consensus is that *T. gondii* disrupts the blood-brain barrier before alterations occur in the CNS. However, questions remain regarding whether tachyzoites directly infect neurons, how the systemic inflammatory environment affects the brain environment through cytokines and inflammatory mediators, evidence of the lytic infection cycle of bradyzoites after cyst formation, and the mechanism of maternal infection. Better research models and extensive research are required to answer these questions.

Distribution of T. gondii Cysts in the Brain Cysts play a major role in the pathogenicity of *T. gondii*. Following infection, *T. gondii* produces cysts in the brain that remain in the human body for a long time,

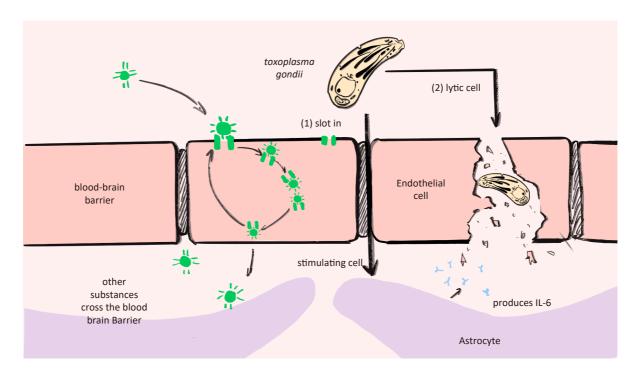


Figure 2. Hypothesis: *Toxoplasma gondii* infection disrupts the blood-brain barrier, causing schizophrenia. Endothelial cells lyse upon the invasion, spread, and proliferation of *T. gondii*. In contrast, when the permeability of the blood-brain barrier increases and restriction decreases, *T. gondii* easily crosses the barrier and enters the brain parenchyma. IL-6, interleukin-6.

causing T. gondii encephalopathy. T. gondii cysts are common in the brain, particularly in the hippocampus and amygdala. Several studies have explored the sites of cyst distribution and their influence on the immune system. Haroon et al. identified neurons as the only cyst-harboring cell population in the CNS^[63]. According to Galvan-Ramirez et al., mice with chronic infections harbor more cysts than animals with acute infections, and the hippocampus and telencephalon are the primary sites of cyst distribution in chronically infected mice^[64]. Barbosa et al. demonstrated through an in vitro model study that T. gondii cysts can disrupt neuronal function, changing the concentration of neurotransmitters in chronically infected mice; encystment also causes enlargement of the neurites and neurite network impairment^[65]. Wu et al. performed a pathological analysis of the distribution of T. gondii cysts in mouse brains and concluded that there was a correlation between the localization and distribution of T. gondii cysts in the brains of infected mice, which displayed dynamic changes over time and induced behavioral neuropsychiatric symptoms during chronic T. gondii infections^[66].

studies However. other have vielded contradictory findings. According to Ingram et al., after early acute infection with T. gondii, mice may permanently lose their intrinsic fear of cats even without long-lasting parasitic cysts or ongoing brain inflammation^[67]. Xiao et al. studied antibody types and found that the distribution of cysts in mouse brains infected with type I T. gondii was directly related to the antibody spectrum, and mice with high levels of the MAG1 antibody showed increased cyst volume and changes in the tendency toward schizophrenia^[68].

In general, it remains controversial whether the distribution and existence of cysts in the brain are necessarily related to schizophrenia, which may also be related to differences in the strains of *T. gondii* and the age of the host.

Structural Changes in the Neural Tissue of the Brain

More studies suggested that the effects of *T. gondii* infection on the nervous system are not limited to the behavioral performance commonly used in the laser swimming test but also involve other underlying mechanisms such as nerve cells and tyrosine hydroxylase. The effects of *T. gondii* infection on neuronal structure and tyrosine hydroxylase expression were investigated by Barbosa et al. (2020). Studies have found that *T. gondii* infection can cause spontaneous cyst

formation in neurons and changes in neuronal structure and tyrosine hydroxylase expression. Specifically, T. gondii infection reduced the length and nuclear size of nerve fibers, the number and total length of dorsal synapses, and the total length and length distribution of neurites. T. gondii infection also significantly reduces the expression of tyrosine hydroxylase. Based on these findings, researchers have suggested that T. gondii infection causes changes in neuronal structure and function and involves the regulation of multiple molecular pathways^[65]. Microglia-neuron interactions and perisomatic inhibitory synapses have also been studied in relation to T. gondii infection. Carrillo et al. showed that microglia and neurons can also become infected with T. gondii, which can cause a loss of peripheral synaptic inhibition, interaction, and contact. Such aberrant neuronal stimulation can cause other diseases. For example, T. gondii infection was found to regulate the interaction between microglia and neurons by activating specific signaling pathways. The study by Carrillo et al. important information for further provides understanding of the mechanisms by which T. gondii affects the nervous system and provides new ideas for the treatment of related diseases [69].

Changes in brain structure may also be linked to immunological responses and neurotransmitters. Chronic inflammatory substances, for example, excite nerve cells, resulting in structural changes, and long-term alterations in neurotransmitters can feedback control synapses and other structures, while the precise mechanisms are unknown, suggesting that when studying intricate mechanisms, it is important to consider the effects of several different factors rather than just one.

Gene Regulation and Epigenetics With the rapid development of epigenetics, many researchers have attempted to explain the association between T. gondii infection and schizophrenia from an epigenetic-regulatory perspective. TEEGR, effector of T. gondii, was reported by Braun et al. to stimulate the EZH2 gene to restrict cell expression, reducing NF-kB signaling and fostering a robust immune response following *T. gondii* infection^[70]. Rovira et al. demonstrated that the catechol-Omethyltransferase (COMT) genotype increases the risk of developing schizophrenia after T. gondii infection; the risk is higher in individuals with a Met/Met phenotype, intermediate in heterozygotes, and lower in individuals with a Val/Val phenotype. It has also been proposed that T. gondii infection decreases COMT activity, increasing dopamine synthesis^[71].

Glutathione S-transferases (GSTs) are a class of isoenzymes encoded by glutathione S-transferase mu-1 (GSTM1), glutathione S-transferases θ -1 and θ -2 (GSTT1 and GSTT2, respectively), glutathione S-transferase pi-1 (GSTP1), and glutathione S-transferase ω -1 (GSTO1). Deleting the entire gene results in insufficient enzyme activity and may cause an increase in the oxidative stress response. Previous studies have linked GST expression to vascular disorders and cancers. Ansari-Lari et al. found an additive effect of T. gondii and the GSTT1-active genotype as risk factors for schizophrenia in an Iranian population. However, further research with larger patient groups is needed to elucidate these findings because this was only a small pilot trial $[^{72}]$.

Epigenetic long noncoding RNAs (IncRNAs) and microRNAs (miRNAs) have been the focus of many studies. These studies have indicated that IncRNA and miRNA expression levels changed dramatically in T. gondii-infected hosts. The sperm of male mice infected with T. gondii showed notable alterations in the miRNA lineage, according to Tyebji et al. Of the total miRNAs mapped (585), 174 were differentially expressed in the Toxoplasma-infected group, with 35 downregulated and 75 upregulated significantly. They showed that compared to non-infected mice, infected mice had higher anxiety and aversion behaviors, with abnormal social interaction patterns that can span generations. In addition, these effects differed between male and female offspring, suggesting that infection in male mice may have caused sex-specific generational changes^[73]. Based on the results of an IncRNA and mRNA co-expression microarray in the brains of mice infected with T. gondii Chinese1 type, Sun et al. revealed that the expression of IncrNA-11496 was downregulated in mice chronically infected with T. gondii, and its dysregulation could further affect cell proliferation, differentiation, and apoptosis by targeting Mef2c^[74]. Wang et al. demonstrated that lncRNA102796 regulates the proliferation of nerve cells by regulating the transcription and expression of oprd1 (a protein that can bind to IncRNA102796) in mouse neuroglial cells (BV-2). These findings indicate that T. gondii-infected nerve cells with downregulated IncRNA102796 expression have an inhibitory effect on nerve cell proliferation, which affects the neurological system^[75]. The results of some studies on epigenetic regulation are summarized in Table 2.

Combination Therapy Outcomes Suggest the Association between Schizophrenia and T. gondii Infection Numerous researchers have discovered correlations between schizophrenia and T. gondii infection in studies of combination and crossmedications for the treatment of both diseases. The findings of several studies support the relationship between T. gondii infection and schizophrenia.

Castano et al. found that sulfadiazine plus pyrimethamine treatment could reverse multiple behavioral and neurocognitive changes in long-term chronic toxoplasmosis by reducing brain cysts and inflammation-related changes, providing a new idea for combination therapy and a basis for the association between schizophrenia and *T. gondii* infections or other diseases^[76]. Valproic acid is a widely used antipsychotic drug, and trimethoprimsulfamethoxazole tablets (TMP-SMZ) are commonly

 Table 2. Current studies regarding epigenetic regulation of Toxoplasma gondii infection

Year	Author	Target	Main findings
2019	Braun et al. ^[70]	The dense granule-resident effector TEEGR	TEEGR binds with the E2F/DP transcription factors to promote <i>EZH2</i> activation, which eventually antagonizes the positive regulation of NF-κB activity by tachyzoites.
2020	Tyebji et al. ^[73]	Sperm small RNA profiles of male mice chronically infected with Toxoplasma gondii	Toxoplasma infection alters small RNA profiles in the F0 sperm and may affect the behavior of F1 and F2 progeny.
2020	Sun et al. ^[74]	The expression of IncRNAs and	The expression of IncRNA-11496 is significantly downregulated and the
2021	Ansari-Lari et al. ^[72]	mRNAs by microarray assay GSTT1 and GSTM1	IncRNA11496/Mef2c/HDAC2 pathway was proposed. An additive effect for <i>T. gondii</i> and <i>GSTT1</i> -active genotype as risk factors for schizophrenia in an Iranian population.
2022	Wang et al. ^[75]	The expression of lncRNA102796 in the brains of mice chronically	The expression of lncRNA102796 was downregulated; lncRNA102796 can regulate the proliferation of nerve cells by regulating the transcription and
2022	Rovira et al. ^[71]	infected with <i>T. gondii</i> COMT Val105/Met158 polymorphism	expression of the <i>oprd1</i> gene. COMT variation and exposure to <i>T. gondii</i> interact and modify the risk for schizophrenia.

Note. GSTT1 encodes glutathione S-transferase T1; GSTM1 encodes glutathione S-transferase M1; COMT, catechol-O-methyltransferase.

used as anti-T. gondii drugs. Using qPCR, Enshaeieh et al. found reduced copy numbers and the transcription level of the bradyzoite-specific protein BAG1, the parasite repetitive DNA fragment REP529, and the highly expressed tachyzoite-specific surface protein SAG1 in the brains of chronically infected mice treated with valproic acid. Additionally, histological analysis of the mouse brain showed that cyst development, lymphocyte perivascular infiltration, and glial nodules were greatly reduced after valproic acid treatment, which showed the same therapeutic level as that in the TMP-SMZ group^[77]. Martynowicz demonstrated is an effective treatment guanabenz neuroinflammation and can significantly reverse the hyperactivity induced by latent toxoplasmosis in C57BL/6 mice. This supports the hypothesis that behavioral disorders in infected hosts may result from dopamine dysfunction or a dysregulation of the GABA pathway^[78].

Changes in Other Metabolic Pathways According to a cross-sectional study conducted by Sagud et al. (2018), for the first time, the ratio of serum triglycerides to high-density lipoprotein cholesterol (HDL-C) was found to be lower in patients with T. gondii antibodies than in seronegative patients. No association was observed between T. gondii seropositivity and serum total cholesterol, lowdensity lipoprotein cholesterol (LDL-C), glucose levels, waist circumference, body mass index, or the MetS ratio. Of note, patients with schizophrenia were taking antipsychotic medications (clozapine, olanzapine, risperidone, and quetiapine) more frequently, which theoretically cause weight gain and metabolic abnormalities, suggesting that T. gondii infection affects lipid metabolism^[79]. In 2022, a survey conducted in East China reached the same conclusion^[80]. In terms of sex hormone metabolism, Borraz-Leon et al. suggested that chronic alterations associated with latent T. gondii infection that affect testosterone feedback regulation may also be risk factors for the development of schizophrenia^[81]. Osman et al. conducted an untargeted metabolomic analysis using liquid chromatography-mass spectrometry to examine plasma metabolomics. They discovered that individuals with schizophrenia or toxoplasmosis had lower levels of xanthine and adenosine monophosphate and higher levels of αhydroxyglutarate than the controls. These results suggest that purine catabolism is a common metabolic alteration in toxoplasmotic infections and schizophrenia^[82]. Since vitamin D deficiency can increase the risk of various chronic mental diseases, Kankova et al. attempted to link vitamin D levels with toxoplasmosis and schizophrenia. However, their findings refute the theory that a decrease in vitamin D levels could be linked to toxoplasmosis^[83].

The function of the metabolites investigated in these studies in the pathophysiology of *T. gondii* infection-related schizophrenia requires more research due to the complexity and heterogeneity of schizophrenia. Further elucidation regarding the underlying pathophysiological mechanisms will assist in clarifying the relationship between *T. gondii* and schizophrenia.

Advances in Research Methods or Animal Models

In vitro-derived Human Neuronal Model Studies

Human nerve cells infected with *T. gondii* can multiply and induce cyst formation, nerve cell death, changes in neural gene expression, and the release of various inflammatory mediators. These findings have been verified using BrainSpheres, a human 3D CNS model derived from human pluripotent stem cells, strengthening the validity of earlier research findings^[84]. A better understanding of host-parasite interactions of *T. gondii* in human neurons and the effects of the parasite on neuronal function is limited by the need for a suitable in vitro model of human neurons^[85]. The advent of cell reprogramming technology, in which somatic cells can be used to generate functional human neurons in vitro, either by using induced pluripotent stem cells (iPSCs) or by directly converting somatic cells into induced neurons, provides an opportunity to develop in vitro human neuronal culture systems. In addition to producing two-dimensional neuronal monolayer cultures that facilitate cellular and molecular mechanistic investigations, induced and iPSC-derived neurons can also be used to create 3D brain organoids, which will aid in disease modeling investigations of the neurological effects of *T. gondii* in the brain.

Human Leukocyte Antigen (HLA) Typing Studies Genema wide association studies (GWAS) has

Genome-wide association studies (GWAS) have regularly linked polymorphisms in various major histocompatibility complex (MHC) regions in schizophrenia. In humans, the MHC region (known as the HLA system) is approximately 40,000 bp long and comprises more than five genes, many of which are involved in the immune system by encoding HLA proteins^[86]. Prior research on the MHC region and schizophrenia, which demonstrated the association between schizophrenia and specific HLA class I and class II alleles, as well as the connection between several autoimmune disorders and schizophrenia,

were confirmed by current GWAS investigations. Polygenic Risk Score (PRS) Lori et al. used the

Psychiatric GWAS Consortium Polygenic Risk Score for Schizophrenia (SCZ-PRS) to assess T. gondiipositive and T. gondii-negative individuals, with schizophrenia diagnosis serving as an outcome variable^[87]. SCZ-PRS assessment does not include MHC variants. Polygenic risk scores (PRS), which assess the genetic liability of several variants into a single individual score, have been used to capture the genetic effects of genome-wide markers that may not individually reach significance in GWAS. Markers in the first training samples were chosen at various p-value threshold levels, and the relevant alleles within each subject were weighted and summed to create scores for the independent replication samples. A significant association between polygenic scores and traits in the replicated samples implies the presence of genetic signals in the selected markers, establishing a common polygenic basis for both datasets. Summary statistics obtained from these datasets are beneficial for PRS analysis because of the polygenic characteristics of schizophrenia and the vast number of samples included in the Schizophrenia Psychiatric Genomics Consortium.

Mouse Models of Gene-environment Interactions

The C-terminus-truncated form of the disrupted-inschizophrenia-1 (DISC1) protein is generated by a translocation associated with several major disorders, including schizophrenia, psychiatric depression, and bipolar disorder. To mimic a possible gene-environment interaction (G × E) model associated with T. gondii-related psychiatric disorders in mice exposed to *T. gondii*, Kannan et al. developed a novel mouse model that induced the expression of DISC1. They used a C-terminustruncated form of the full-length protein as a dominant-negative molecular tool (DN-DISC1) to alter the expression of endogenous full-length DISC1 to elucidate the role of DISC1. In this geneenvironment interaction model, T. gondii exposure caused DN-DISC1-expressing mice to exhibit sexdependent prepulse suppression of acoustic startle^[88]. This provides a new experimental system and scheme for future research.

Conclusion

In addition to uncovering novel therapeutic targets and diagnostic indicators, research on T. gondii infection and schizophrenia contributes to our understanding of the pathophysiology of Mental disease. It also offers a pertinent theoretical framework for forensic psychology, sociology, psychiatry, and other applicable fields. Many studies have demonstrated a strong association between T. gondii infection and schizophrenia, supported by serological and animal experimental data. Many researchers have attempted to explore the association between *T. gondii* infection schizophrenia in terms of pathological anatomy, imaging, behavior, immune molecules, and genes. Their findings demonstrate that researchers need to continue studying this direction, especially the underlying mechanisms at the genetic and epigenetic levels. It is important to note that schizophrenia is a complex illness. While numerous studies have demonstrated the null hypothesis of a relationship, only a small percentage have refuted the theory that T. gondii infection causes schizophrenia. Future studies should pay more attention to related factors or confounders, such as the course of acute or chronic T. gondii infection, whether a patient is in the stable phase or experiencing an episode of schizophrenia during the statistical investigation, age, sex, drug use, and the type of subject used in animal experiments. In particular, researchers should pay attention to the different strains of *T. gondii* that caused the infection because the virulence and tendency of *T. gondii* may have important effects on the experimental results. For example, the type I strain is associated with mental illness^[5,6]. Furthermore, the dominant strain is dispersed throughout many regions; for example, the dominant strain in Chinese1 was found in China^[7]. Competing Interests The authors declare that they

have no competing interests.

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REFERENCES

- 1. Montoya JG, Liesenfeld O. Toxoplasmosis. Lancet, 2004; 363, 1965-76.
- 2. Nicolle C, Manceaux L. Sur une infection à corps de Leishman (ou organismes voisins) du gondi. C R Seances Acad Sci, 1908; 147. 763-6.
- 3. Splendore A. Un nuovo protozoa parassita de' conigli. Incontrato nelle lesioni anatomiche d'une malattia che ricorda in molti punti il Kala-azar dell' uomo. Nota preliminare Pel Rev

- Soc Scient Sao Paulo, 1908; 3, 109-12.
- Howe DK, Sibley LD. *Toxoplasma gondii* comprises three clonal lineages: correlation of parasite genotype with human disease. J Infect Dis, 1995; 172, 1561–6.
- Xiao JC, Jones-Brando L, Talbot CC Jr, et al. Differential effects of three canonical *Toxoplasma* strains on gene expression in human neuroepithelial cells. Infect Immun, 2011; 79, 1363–73.
- Groër MW, Yolken RH, Xiao JC, et al. Prenatal depression and anxiety in *Toxoplasma gondii*-positive women. Am J Obstet Gynecol, 2011; 204, 433.e1–7.
- Chaichan P, Mercier A, Galal L, et al. Geographical distribution of *Toxoplasma gondii* genotypes in Asia: a link with neighboring continents. Infect Genet Evol, 2017; 53, 227–38.
- 8. Shen JL, Yu L. Prevalence and fundamental researches of prevention and treatment of toxoplasmosis in China: an overview. Chin J Schisto Control, 2019; 31, 71–6. (In Chinese)
- Graham AK, Fong C, Naqvi A, et al. Toxoplasmosis of the central nervous system: manifestations vary with immune responses. J Neurol Sci, 2021; 420, 117223.
- Luft BJ, Remington JS. Toxoplasmic encephalitis in AIDS. Clin Infect Dis, 1992; 15, 211–22.
- Maldonado YA, Read JS, Committee on Infectious Diseases. Diagnosis, treatment, and prevention of congenital toxoplasmosis in the united states. Pediatrics, 2017; 139, e20163860.
- Luft BJ, Conley F, Remington JS, et al. Outbreak of centralnervous-system toxoplasmosis in western Europe and North America. Lancet. 1983: 321. 781–4.
- Suzuki Y, Yang Q, Yang S, et al. IL-4 is protective against development of toxoplasmic encephalitis. J Immunol, 1996; 157, 2564–9.
- 14. Yao Y, Shi TY, Shu PY, et al. *Toxoplasma gondii* infection and brain inflammation: a two-sample mendelian randomization analysis. Heliyon, 2024; 10, e24228.
- Lüder CGK, Giraldo-Velásquez M, Sendtner M, et al. Toxoplasma gondii in primary rat CNS cells: differential contribution of neurons, astrocytes, and microglial cells for the intracerebral development and stage differentiation. Exp Parasitol, 1999; 93, 23–32.
- Olariu TR, Remington JS, McLeod R, et al. Severe congenital toxoplasmosis in the United States: clinical and serologic findings in untreated infants. Pediatr Infect Dis J, 2011; 30, 1056-61.
- 17. Chen XJ, Chen B, Hou XQ, et al. Association between *Toxoplasma gondii* infection and psychiatric disorders in Zhejiang, Southeastern China. Acta Trop, 2019; 192, 82–6.
- Veleva I, Stoychev K, Stoimenova-Popova M, et al. *Toxoplasma gondii* seropositivity and cognitive function in adults with schizophrenia. Schizophr Res Cogn, 2022; 30, 100269.
- Grada S, Mihu AG, Petrescu C, et al. *Toxoplasma gondii* Infection in patients with psychiatric disorders from western Romania. Medicina, 2022; 58, 208.
- Liu TX, Gao P, Bu DY, et al. Association between *Toxoplasma gondii* infection and psychiatric disorders: a cross-sectional study in China. Sci Rep, 2022; 12, 15092.
- 21. Mao FZ, Yang YG, Chen YY, et al. Seroprevalence and risk factors of *Toxoplasma gondii* infection among high-risk populations in Jiangsu Province, Eastern China. Front Cell Infect Microbiol, 2021; 11, 783654.
- 22. Ademe M, Kebede T, Teferra S, et al. Is latent *Toxoplasma gondii* infection associated with the occurrence of schizophrenia? A case-control study. PLoS One, 2022; 17, e0270377
- 23. Contopoulos-Ioannidis DG, Gianniki M, Ai-Nhi Truong A, et al. Toxoplasmosis and schizophrenia: a systematic review and meta-analysis of prevalence and associations and future

- directions. Psychiatr Res Clin Pract, 2022; 4, 48-60.
- 24. Daré LO, Bruand PE, Gérard D, et al. Associations of mental disorders and neurotropic parasitic diseases: a meta-analysis in developing and emerging countries. BMC Public Health, 2019; 19, 1645.
- 25. Sutterland AL, Kuin A, Kuiper B, et al. Driving us mad: the association of *Toxoplasma gondii* with suicide attempts and traffic accidents a systematic review and meta-analysis. Psychol Med, 2019; 49, 1608–23.
- 26. Sutterland AL, Fond G, Kuin A, et al. Beyond the association. Toxoplasma gondii in schizophrenia, bipolar disorder, and addiction: systematic review and meta-analysis. Acta Psychiatr Scand, 2015; 132, 161–79.
- Monroe JM, Buckley PF, Miller BJ. Meta-analysis of anti-Toxoplasma gondii IgM antibodies in acute psychosis. Schizophr Bull, 2015; 41, 989–98.
- Montazeri M, Moradi E, Moosazadeh M, et al. Relationship between *Toxoplasma gondii* infection and psychiatric disorders in Iran: a systematic review with meta-analysis. PLoS One, 2023; 18, e0284954.
- Piekarski G, Zippelius HM, Witting PA. Auswirkungen einer latenten *Toxoplasma*-infektion auf das lernvermögen von weissen laboratoriumsratten und -mäusen. Z Parasitenkd, 1978; 57, 1–15.
- 30. Daniels BP, Sestito SR, Rouse ST. An expanded task battery in the Morris water maze reveals effects of *Toxoplasma gondii* infection on learning and memory in rats. Parasitol Int, 2015; 64, 5–12.
- Vyas A, Kim SK, Giacomini N, et al. Behavioral changes induced by *Toxoplasma* infection of rodents are highly specific to aversion of cat odors. Proc Natl Acad Sci USA, 2007; 104, 6442-7.
- 32. Boillat M, Hammoudi PM, Dogga SK, et al. Neuroinflammationassociated aspecific manipulation of mouse predator fear by *Toxoplasma qondii*. Cell Rep, 2020; 30, 320-34. e6.
- Webster JP, Kaushik M, Bristow GC, et al. *Toxoplasma gondii* infection, from predation to schizophrenia: can animal behaviour help us understand human behaviour? J Exp Biol, 2013; 216. 99-112.
- 34. Holub D, Flegr J, Dragomirecká E, et al. Differences in onset of disease and severity of psychopathology between toxoplasmosis-related and toxoplasmosis-unrelated schizophrenia. Acta Psychiatr Scand, 2013; 127, 227–38.
- 35. Sutterland AL, Mounir DA, Ribbens JJ, et al. *Toxoplasma gondii* infection and clinical characteristics of patients with schizophrenia: a systematic review and meta-analysis. Schizophr Bull Open, 2020; 1, sgaa042.
- 36. Brüne M. Schizophrenia as parasitic behavior manipulation: can we put together the pieces of an evolutionary puzzle? World Psychiatry, 2020; 19, 119-20.
- 37. Stibbs HH. Changes in brain concentrations of catecholamines and indoleamines in *Toxoplasma gondii* infected mice. Ann Trop Med Parasitol, 1985; 79, 153–7.
- Ibrahim Ali M, Abdel Gawad Mousa Ismail M, Abd-Elftah Abd-Allah G, et al. Toxoplasmosis in schizophrenic patients: immune-diagnosis and serum dopamine level. Pak J Biol Sci, 2020; 23, 1131–7.
- Omidian M, Asgari Q, Bahreini MS, et al. Acute toxoplasmosis can increase serum dopamine level. J Parasit Dis, 2022; 46, 337–42
- Prandovszky E, Gaskell E, Martin H, et al. The neurotropic parasite *Toxoplasma gondii* increases dopamine metabolism. PLoS One, 2011; 6, e23866.
- Gaskell EA, Smith JE, Pinney JW, et al. A unique dual activity amino acid hydroxylase in *Toxoplasma gondii*. PLoS One, 2009; 4, e4801.

- 42. McFarland R, Wang ZT, Jouroukhin Y, et al. *AAH2* gene is not required for dopamine-dependent neurochemical and behavioral abnormalities produced by *Toxoplasma* infection in mouse. Behav Brain Res, 2018; 347, 193–200.
- Afonso C, Paixão VB, Klaus A, et al. *Toxoplasma*-induced changes in host risk behaviour are independent of parasitederived AaaH2 tyrosine hydroxylase. Sci Rep, 2017; 7, 13822.
- 44. Strobl JS, Goodwin DG, Rzigalinski BA, et al. Dopamine stimulates propagation of *Toxoplasma gondii* tachyzoites in human fibroblast and primary neonatal rat astrocyte cell cultures. J Parasitol, 2012; 98, 1296–9.
- David CN, Frias ES, Szu JI, et al. GLT-1-dependent disruption of CNS glutamate homeostasis and neuronal function by the protozoan parasite *Toxoplasma gondii*. PLoS Pathog, 2016; 12, e1005643.
- 46. Acquarone M, Poleto A, Perozzo AF, et al. Social preference is maintained in mice with impaired startle reflex and glutamate/D-serine imbalance induced by chronic cerebral toxoplasmosis. Sci Rep, 2021; 11, 14029.
- 47. Lucchese G. From toxoplasmosis to schizophrenia via NMDA dysfunction: peptide overlap between *Toxoplasma gondii* and *N*-methyl-D-aspartate receptors as a potential mechanistic link. Front Psychiatry, 2017; 8, 37.
- Brooks JM, Carrillo GL, Su JM, et al. Toxoplasma gondii infections alter GABAergic synapses and signaling in the central nervous system. mBio, 2015; 6, e01428–15.
- Pearce BD, Massa N, Goldsmith DR, et al. *Toxoplasma gondii* effects on the relationship of kynurenine pathway metabolites to acoustic startle latency in schizophrenia vs. control subjects. Front Psychiatry, 2020; 11, 552743.
- Mahmoud ME, Ihara F, Fereig RM, et al. Induction of depression-related behaviors by reactivation of chronic Toxoplasma gondii infection in mice. Behav Brain Res, 2016; 298. 125–33.
- 51. Konradt C, Ueno N, Christian DA, et al. Endothelial cells are a replicative niche for entry of *Toxoplasma gondii* to the central nervous system. Nat Microbiol, 2016; 1, 16001.
- 52. Kim H, Hong SH, Jeong HE, et al. Microfluidic model for in vitro acute *Toxoplasma gondii* infection and transendothelial migration. Sci Rep, 2022; 12, 11449.
- Estato V, Stipursky J, Gomes F, et al. The neurotropic parasite Toxoplasma gondii induces sustained neuroinflammation with microvascular dysfunction in infected mice. Am J Pathol, 2018; 188. 2674–87.
- 54. Hamdani N, Bengoufa D, Godin O, et al. Immunoglobulin subclass distribution in bipolar disorder and schizophrenia: potential relationship with latent *Toxoplasma Gondii* infection. BMC Psychiatry, 2018; 18, 239.
- 55. Fond G, Boyer L, Schürhoff F, et al. Latent toxoplasma infection in real-world schizophrenia: Results from the national FACE-SZ cohort. Schizophr Res, 2018; 201, 373–80.
- 56. Marcos AC, Siqueira M, Alvarez-Rosa L, et al. *Toxoplasma gondii* infection impairs radial glia differentiation and its potential to modulate brain microvascular endothelial cell function in the cerebral cortex. Microvasc Res, 2020; 131, 104024
- 57. Schneider CA, Figueroa Velez DX, Orchanian SB, et al. Toxoplasma gondii dissemination in the brain is facilitated by infiltrating peripheral immune cells. mBio, 2022; 13, e0283822.
- 58. Castaño Barrios L, Da Silva Pinheiro AP, Gibaldi D, et al. Behavioral alterations in long-term *Toxoplasma gondii* infection of C57BL/6 mice are associated with neuroinflammation and disruption of the blood brain barrier. PLoS One, 2021; 16, e0258199.
- 59. Carrillo GL, Su JM, Cawley ML, et al. Complement-dependent

- loss of inhibitory synapses on pyramidal neurons following *Toxoplasma gondii* infection. J Neurochem, 2023.
- 60. Xiao JC, Li Y, Rowley T, et al. Immunotherapy targeting the PD-1 pathway alleviates neuroinflammation caused by chronic *Toxoplasma* infection. Sci Rep, 2023; 13, 1288.
- 61. Romero Núñez E, Blanco Ayala T, Vázquez Cervantes GI, et al. Pregestational exposure to *T. gondii* produces maternal antibodies that recognize fetal brain mimotopes and induces neurochemical and behavioral dysfunction in the offspring. Cells. 2022: 11. 3819.
- Bi XL, Fu XY, Xue S, et al. Expression of CD47 and its ligands in pregnant mice infected with *Toxoplasma gondii* during pregnancy. Chin J Schistosomiasis Control, 2023; 35, 51–62. (In Chinese)
- 63. Haroon F, Händel U, Angenstein F, et al. *Toxoplasma gondii* actively inhibits neuronal function in chronically infected mice. PLoS One, 2012; 7, e35516.
- 64. de la Luz Galván-Ramírez M, Salas-Lais AG, Dueñas-Jiménez SH, et al. Kinematic locomotion changes in C57BL/6 mice infected with *Toxoplasma* strain ME49. Microorganisms, 2019; 7, 573.
- 65. Barbosa JL, Béla SR, Ricci MF, et al. Spontaneous *T. gondii* neuronal encystment induces structural neuritic network impairment associated with changes of tyrosine hydroxilase expression. Neurosci Lett, 2020; 718, 134721.
- 66. Wu MY, Cai R, Li YF, et al. Dynamic pathological changes of Toxoplasma cysts in mouse brain during chronic infection. Acta Univ Med Anhui, 2023; 58, 184–8. (In Chinese)
- 67. Ingram WM, Goodrich LM, Robey EA, et al. Mice infected with low-virulence strains of *Toxoplasma gondii* lose their innate aversion to cat urine, even after extensive parasite clearance. PLoS One. 2013: 8, e75246.
- 68. Xiao JC, Li Y, Prandovszky E, et al. Behavioral abnormalities in a mouse model of chronic toxoplasmosis are associated with MAG1 antibody levels and cyst burden. PLoS Negl Trop Dis, 2016; 10, e0004674.
- 69. Carrillo GL, Ballard VA, Glausen T, et al. *Toxoplasma* infection induces microglia-neuron contact and the loss of perisomatic inhibitory synapses. Glia, 2020; 68, 1968–86.
- Braun L, Brenier-Pinchart MP, Hammoudi PM, et al. The Toxoplasma effector TEEGR promotes parasite persistence by modulating NF-κB signalling via EZH2. Nat Microbiol, 2019; 4, 1208–20.
- 71. Rovira P, Gutiérrez B, Sorlózano-Puerto A, et al. *Toxoplasma gondii* seropositivity interacts with catechol-*O*-methyltransferase Val105/158Met variation increasing the risk of schizophrenia. Genes, 2022; 13, 1088.
- Ansari-Lari M, Zendehboodi Z, Masoudian M, et al. Additive effect of glutathione S-transferase T1 active genotype and infection with *Toxoplasma gondii* for increasing the risk of schizophrenia. Nord J Psychiatry, 2021; 75, 275–80.
- Tyebji S, Hannan AJ, Tonkin CJ. Pathogenic infection in male mice changes sperm small RNA profiles and transgenerationally alters offspring behavior. Cell Rep, 2020; 31, 107573.
- 74. Sun XH, Wang T, Wang YL, et al. Downregulation of IncRNA-11496 in the brain contributes to microglia apoptosis via regulation of Mef2c in chronic *T. gondii* infection mice. Front Mol Neurosci, 2020; 13, 77.
- Wang ZX, Xiong SS, Sun XH, et al. Differential expression and action mechanism of IncRNA102796 in the brain of mice with chronic infection of *Toxoplasma gondii*. Chin J Parasitol Parasit Dis, 2022; 40, 187–93. (In Chinese)
- 76. Castaño BL, Silva AA, Hernandez-Velasco LL, et al. Sulfadiazine plus pyrimethamine therapy reversed multiple behavioral and neurocognitive changes in long-term chronic toxoplasmosis by

- reducing brain cyst load and inflammation-related alterations. Front Immunol, 2022; 13, 822567.
- Enshaeieh M, Saadatnia G, Babaie J, et al. Valproic acid inhibits chronic *Toxoplasma* infection and associated brain inflammation in mice. Antimicrob Agents Chemother, 2021; 65, e0100321.
- 78. Martynowicz J, Augusto L, Wek RC, et al. Guanabenz reverses a key behavioral change caused by latent toxoplasmosis in mice by reducing neuroinflammation. mBio, 2019; 10, e00381–19.
- Sagud M, Vlatkovic S, Svob Strac D, et al. Latent *Toxoplasma gondii* infection is associated with decreased serum triglyceride to high-density lipoprotein cholesterol ratio in male patients with schizophrenia. Compr Psychiatry, 2018; 82, 115–20.
- 80. Xu F, Ma XY, Zhu YW, et al. Effects of *Toxoplasma gondii* infection and schizophrenia comorbidity on serum lipid profile: a population retrospective study from Eastern China. Microb Pathog, 2020; 149, 104587.
- 81. Borráz-León JI, Rantala MJ, Luoto S, et al. *Toxoplasma gondii* and psychopathology: latent infection is associated with interpersonal sensitivity, psychoticism, and higher testosterone levels in men, but not in women. Adapt Human Behav Physiol, 2021; 7, 28–42.
- 82. Osman E, Mohammad Zahariluddin AS, Sharip S, et al.

- Metabolomic profiling reveals common metabolic alterations in plasma of patients with *Toxoplasma* infection and schizophrenia. Genes, 2022; 13, 1482.
- 83. Kankova S, Bicikova M, Macova L, et al. Latent toxoplasmosis and vitamin D concentration in humans: three observational studies. Folia Parasitol, 2021; 68, 2021.005.
- 84. Correa Leite PE, de Araujo Portes J, Pereira MR, et al. Morphological and biochemical repercussions of *Toxoplasma gondii* infection in a 3D human brain neurospheres model. Brain Behav Immun Health, 2020; 11, 100190.
- 85. Halonen SK. Use of *in vitro* derived human neuronal models to study host-parasite interactions of *Toxoplasma gondii* in neurons and neuropathogenesis of chronic toxoplasmosis. Front Cell Infect Microbiol, 2023; 13, 1129451.
- 86. Parks S, Avramopoulos D, Mulle J, et al. HLA typing using genome wide data reveals susceptibility types for infections in a psychiatric disease enriched sample. Brain Behav Immun, 2018; 70, 203–13.
- 87. Lori A, Avramopoulos D, Wang AW, et al. Polygenic risk scores differentiate schizophrenia patients with *Toxoplasma gondii* compared to *toxoplasma* seronegative patients. Compr Psychiatry, 2021; 107, 152236.
- 88. Kannan G, Prandovszky E, Severance E, et al. A new *T. gondii* mouse model of gene-environment interaction relevant to psychiatric disease. Scientifica, 2018; 2018, 7590958.